

Review Article

Sulforaphane: Its “Coming of Age” as a Clinically Relevant Nutraceutical in the Prevention and Treatment of Chronic Disease

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A growing awareness of the mechanisms by which phytochemicals can influence upstream endogenous cellular defence processes has led to intensified research into their potential relevance in the prevention and treatment of disease. Pharmaceutical medicine has historically looked to plants as sources of the starting materials for drug development; however, the focus of nutraceutical medicine is to retain the plant bioactive in as close to its native state as possible. As a consequence, the potency of a nutraceutical concentrate or an extract may be lower than required for significant gene expression. The molecular structure of bioactive phytochemicals to a large extent determines the molecule's bioavailability. Polyphenols are abundant in dietary phytochemicals, and extensive *in vitro* research has established many of the signalling mechanisms involved in favourably modulating human biochemical pathways. Such pathways are associated with core processes such as redox modulation and immune modulation for infection control and for downregulating the synthesis of inflammatory cytokines. Although the relationship between oxidative stress and chronic disease continues to be affirmed, direct-acting antioxidants such as vitamins A, C, and E, beta-carotene, and others have not yielded the expected preventive or therapeutic responses, even though several large meta-analyses have sought to evaluate the potential benefit of such supplements. Because polyphenols exhibit poor bioavailability, few of their impressive *in vitro* findings have been replicated *in vivo*. SFN, an aliphatic isothiocyanate, emerges as a phytochemical with comparatively high bioavailability. A number of clinical trials have demonstrated its ability to produce favourable outcomes in conditions for which there are few satisfactory pharmaceutical solutions, foreshadowing the potential for SFN as a clinically relevant nutraceutical. Although myrosinase-inert broccoli sprout extracts are widely available, there now exist myrosinase-active broccoli sprout supplements that yield sufficient SFN to match the doses used in clinical trials.

1. Introduction

We live in an era where modern medicine is strongly focused on relief of symptoms with pharmaceuticals, providing many solutions to address this demand. It is becoming increasingly apparent, however, that for the diseases which cause most distress at the individual level, pharmaceuticals typically provide only short-lived symptomatic relief. Few if any modern pharmaceuticals modulate fundamental etiological disease processes.

As a consequence, there is a groundswell of interest in phytochemical solutions which may potentially target the fundamental *upstream* causes of disease [1, 2]. Plant-derived

bioactive compounds are already emerging as candidate molecules with significant therapeutic potential in human health [3]. Numerous mechanistic investigations of phytochemical bioactives are already helping to elucidate the pathophysiology of both chronic diseases and acute self-limiting conditions [4]. It is generally considered that such findings may inform the development of new therapeutic solutions. Although pharmaceutical medicine has historically looked to plants as sources of the starting materials for drug development, the ultimate therapeutic molecule is typically quite different from the original plant-derived source. By contrast, the focus of nutraceutical medicines is to retain the plant bioactive in as close to its native state as possible. The challenge for

TABLE 1: Major randomized placebo-controlled trials investigating the effects of the antioxidant supplement on prevention of diabetes or glucose homeostasis.

Study	Study population	Duration (years)	Antioxidants (daily dose)	Endpoint	Results
Women's Health Study	38,716 healthy U.S. women	10	Vitamin E (α -tocopherol: 600 IU; 933.3 μ mol)	Incident diabetes	No effect
Women's Antioxidant Cardiovascular Study	6,574 nondiabetic U.S. women at high risk of cardiovascular disease	9.2	Vitamin E (α -tocopherol: 300 IU; 466.7 μ mol) Vitamin C (500 mg; 2.84 Mmol) Beta-carotene (25 mg; 46.6 μ mol)	Incident diabetes	No effect
Physician Health Study	22,071 healthy U.S. male physicians	12	Beta-carotene (25 mg; 46.6 μ mol)	Incident diabetes	No effect
Alpha-Tocopherol, Beta-Carotene Cancer Prevention Study	27,379 nondiabetic male Finnish smokers	12.5	Vitamin E (α -tocopherol 50 mg; 116.1 μ mol) Beta-carotene (20 mg; 37.3 μ mol)	Incident diabetes	No effect
Supplementation with Antioxidant Vitamins and Minerals study	3,146 nondiabetic French	7.5	Vitamin C (120 mg; 681.4 μ mol) Vitamin E (30 mg; 104.5 μ mol) Beta-carotene (6 mg; 11.2 μ mol) Selenium (100 μ g; 1.27 μ mol) Zinc (20 mg; 306 μ mol)	Fasting glucose	No effect

developers of nutraceutical supplements is that the potency of such nutraceutical concentrates or extracts may be below the threshold required to nutrigenomically induce the gene expression required for a significant therapeutic response.

1.1. Searching for Upstream Factors. Because homeostasis in human cells is reliant on the dynamic integration of many core biochemical processes, a search for *upstream* factors in the etiological processes of disease is the focus of considerable global research; such research is closely focused on investigating signalling pathways within cells and organelles. Prior to the introduction of better hygiene practices, the global disease burden was dominated by infectious diseases. By contrast, more recent decades have seen a steady increase in levels of morbidity and mortality rates from chronic disease, justifying the claim that chronic disease has reached epidemic proportions [5].

As one example, the increasing global prevalence of cardiovascular disease (CVD) and type 2 diabetes (T2DM) is dominant in the current trajectory for chronic disease. It is emerging [6] that the primary upstream factor which links endothelial dysfunction with CVD and T2DM and described as cardiometabolic disease is closely related to oxidative stress [6–11].

More recently, mechanistic studies link cardiometabolic dysfunction with intestinal dysfunction and subsequent metabolic endotoxaemia. The cell walls of gram-negative bacteria increase luminal levels of lipopolysaccharides (LPS) that are detected by and bind to Toll-like receptor 4 (TLR4). This initiates the activation of Nf- κ B with the subsequent generation of inflammatory cytokines that are systemically absorbed [12]. At least three apparently distinct mechanisms—endoplasmic reticulum stress, toll-like receptor (TLR) 4 activation, and changes in gut microbiota—have been identified

as triggers of obesity-associated metabolic inflammation [13]. SFN, the focus of this review, has been identified as a molecule that can reduce inflammation via inhibition of LPS-TLR4 binding [14, 15]; this mechanism is further discussed in Section 7.5.

1.2. Failed Antioxidant Trials. Attempts to use the classical antioxidant vitamins to enhance endothelial function and related glucose modulation have largely resulted in no response in some studies and adverse effects in others [9, 16–19].

A 2010 meta-analysis [20] of major randomized placebo-controlled trials (98,886 subjects in total, Table 1) investigating the effects of the antioxidant supplement on prevention of diabetes or effect on glucose homeostasis showed no effect from vitamin E, vitamin C, beta-carotene, selenium, zinc, and combinations of these.

Similar meta-analyses also fail to demonstrate significant chemoprotection or preventive benefits against cancer and cardiovascular disease via antioxidant vitamins [16, 21–23]. These findings suggest the possibility that intervention with phytochemicals as redox-modulating biomolecules might provide an alternative but effective strategy.

1.3. SFN and Type 2 Diabetes Intervention Trials. Whereas Table 1 lists large-scale clinical trials considering T2DM risk in thousands of individuals over long periods, the studies in which SFN has been utilised as the intervention material are few, are of short duration, and include small numbers of participants.

To query whether SFN as an indirect antioxidant could modify disease risk in T2DM where direct-acting antioxidants seemed unable, a 4-week randomized controlled clinical trial [24] was conducted in 2011 to investigate the effect of

5 grams (yielding 112.5 μmol SFN) and 10 grams (yielding 225 μmol SFN) daily of broccoli sprout powder on 81 T2DM patients and using cardiometabolic biomarkers as the outcome measures. The results provided the first data to show that SFN could reduce lipid peroxidation, especially significant at the higher dose. In particular, favourable redox status was demonstrated by a decrease in plasma malondialdehyde (MDA) and oxidised LDL (OX-LDL). As increased lipid peroxidation in diabetes has been implicated as an important factor in the pathogenesis of T2DM complications, the researchers considered the potential for SFN to play a role in the prevention of T2DM and its secondary effects.

In general, the studies which have used SFN-yielding broccoli sprouts or supplements to enhance cellular defences have shown promising mechanistic findings but inconsistent clinical responses [25–30]. A 2018 study investigating biomarkers of inflammation in overweight but otherwise healthy adults showed significant downregulation of two such biomarkers; this is expanded in Section 7.6 with the clinical trial data discussed in Section 7.7. These data hold promise for the clinical application of SFN in inflammation-related conditions.

1.4. The Origin of Antioxidant Supplements as Therapy. The notion that aging was due to a state of oxidative stress within cells emerged in the 1950s from Dr. Denham Harman, a research chemist in the petrochemical industry who subsequently joined the faculty of the University of Nebraska Medical Center. His “free radical theory of aging” postulates that the typical changes that occur during aging are caused by free radical reactions [31].

The theory gained initial support by others including Nobel laureate, Linus Pauling whose hypotheses on ascorbic acid deficiency as an etiological factor in cancer and acute infectious illnesses earned him enormous popularity with consumers but derision within much of the scientific community [32]. It could be argued that the promotion of vitamin C as a “cure” for the common cold and for cancer heralded the onset of a huge upsurge in sales of antioxidant vitamins.

It would seem that because plant-based diets had been shown epidemiologically to be protective against a range of diseases [33], it had been erroneously assumed that the protective effect was conferred by the presence of vitamins like A, C and E and beta-carotene. Had these early researchers also considered that plant foods are endowed with an extensive range of bioactive phytochemicals functioning via different mechanisms, they may not have drawn this conclusion [34–36].

1.5. Addressing a More Nuanced View of Redox Balance. The study of the relationship between oxidative stress, aging, and disease remains popular, with investigators striving to identify interventions that are capable of modulating the disease-causing processes [37]. The free radical-antioxidant theory proposed decades ago proved to be too simplistic; more recent research reveals multiple signalling processes at play [8]. As we observe the unfolding of the complex relationships governing endogenous cellular mechanisms, a close interconnectedness between redox balance, inflammation,

and endoplasmic reticulum stress emerges [38]. What this indicates is that any therapeutic attempt to successfully intervene must either address each process individually or intervene sufficiently *upstream* at a point that can beneficially influence multiple *downstream* targets.

It is within this framework of redox-associated disease that this review considers mechanisms by which the Brassica-derived phytochemical, sulforaphane (SFN), may be utilised therapeutically to modulate the *upstream* cellular perturbations that contribute to the etiology of disease.

A number of large systematic reviews and meta-analyses, including Cochrane Reviews, have concluded that although oxidative stress underpins common chronic diseases, antioxidant vitamins do not lead to reduction in disease risk [16, 22, 39–46].

2. Phytochemicals as Inducers of Endogenous Defences

A possible alternative approach to the modulation of oxidative stress by direct-acting antioxidant vitamins involves the application of phytochemicals with nutrigenomic potential [47]. By definition, a phytochemical is a plant-derived chemical substance that is biologically active but typically nonnutritive [48]; nutrigenomics describes the way in which phytochemicals and nutrients may affect gene expression. As such, the application of nutrigenomic principles may allow effective dietary intervention strategies to recover normal homeostasis and to prevent or even treat diet-related diseases [49]. Phytochemicals are abundant in the food supply and have been classified according to their molecular structure, a property which contributes to their observed beneficial on human health [36, 50].

2.1. Polyphenols: Their Clinical Potential. Polyphenolic bioactives derived from plant species have been extensively researched in relation to their mechanisms of action in human cells and for their clinical potential in modulating disease-causing processes [51]. Such molecules have significant *in vitro* antioxidant potential, but their low bioavailability [52, 53] limits their clinical usefulness as systemic antioxidants.

Consequently, although extensively studied *in vitro*, attempts to replicate these findings *in vivo* have been disappointing and it is generally considered that the large molecular weight and structure of these molecules is a significant factor impeding their bioavailability [50, 54]. Another role for polyphenols has more recently been identified in relation to their prebiotic and other beneficial effects on the gut microbiota [55].

A very recent meta-analysis of clinical trials in which polyphenol-based supplements were evaluated for their beneficial effects on specific markers of cardiovascular risk and cognitive status concluded that definitive recommendations for the use of these compounds could not yet be made and that additional characterisation of pharmacokinetics and safety is required [56]. The bioavailability of the polyphenolic phytochemicals so abundant in the food supply lies between 1% and 10%. This is discussed in some depth in Section 6 of an earlier review by this author [54].

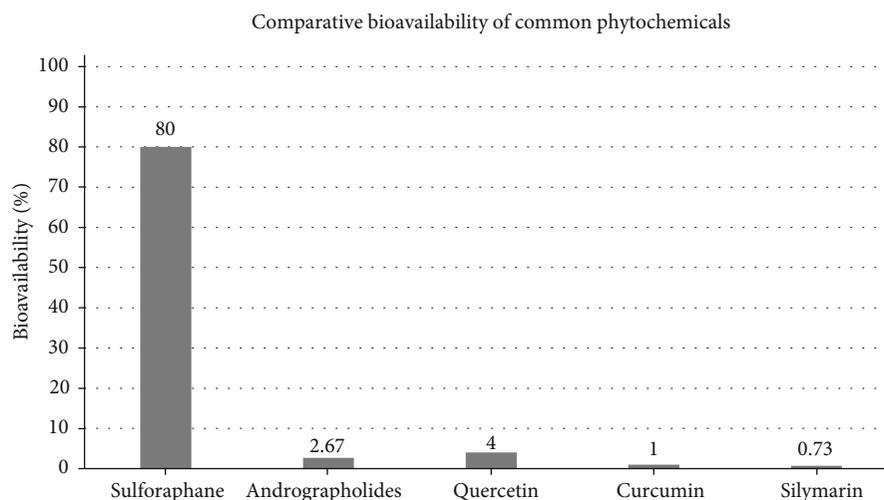


FIGURE 1: Comparative bioavailability of phytochemicals commonly used in dietary supplements (appears as Figure 3 in [54]).

SFN, derived primarily from broccoli and with absolute bioavailability of around 80% [57], shows promise as a nutrigenomically active compound capable of increasing several endogenous antioxidant compounds via the transcription factor, Nrf2 [58]. SFN, an aliphatic isothiocyanate [59], emerges as a phytochemical with comparatively high bioavailability due to its low molecular weight and log P value of 0.23 [60] when evaluated using the commonly used octan-1-ol and water system and where log $P = 0$ represents equal partitioning between the solvent and water and where a value > 0 represents a higher concentration in the lipid phase [61]. Figure 1 highlights the differences in bioavailability when comparing SFN with polyphenols commonly used in dietary supplements.

2.2. The Nutrigenomic Link to Endogenous Antioxidants.

Nutrigenomically active phytochemicals exhibit a number of different mechanisms as modulators of the expression of genes coding for enzymes which are active in diverse pathways [62]. One of the intended effects of this strategy is to increase the production of *endogenous* antioxidant compounds including the antioxidant enzymes. Whilst some phytochemicals may upregulate cellular endogenous defences, others may downregulate pathways associated with undesirable prolonged inflammation. The key transcription factors responsible for the induction of redox-modulating and inflammation-promoting gene expression are, respectively, Nrf2 and NK- κ B; these transcription factors act both independently and cooperatively via cross talk that is not yet fully understood [63].

Although such plant-derived compounds may exhibit direct antioxidant activity, it is their *indirect* antioxidant effect which is of most interest, due to the catalytic effect of the antioxidant enzymes in quenching reactive oxygen and reactive nitrogen species (ROS and RNS) compared with nonenzyme antioxidants which exhibit only a one-for-one stoichiometric effect [64, 65]. There is considerable evidence to show that induction of such cytoprotective compounds has multiple beneficial effects [66–69].

Germinated broccoli seeds yield a nutrigenomically active isothiocyanate, SFN; this review focuses on the properties of SFN as they relate to its antioxidant, anti-inflammatory, and antimicrobial effects. Furthermore, this discussion reviews the doses used in relevant clinical trials with a view to evaluating whether these doses are practical for SFN to be considered as a nutraceutical with broad clinical application and whether it may be considered as an efficacious nutraceutical in the prevention and treatment of chronic disease.

2.3. Determining Clinical Potential of a Phytochemical. When considering the likelihood that a particular phytochemical may exhibit clinical potential, two important factors bear mention. Firstly, the bioactive molecule must have sufficient potency to induce adequate gene expression in the target gene or genes; secondly, the bioactive must be sufficiently bioavailable that the concentration measurable in the bloodstream or target tissue is able to match the concentrations measured in the *in vivo* studies for which gene expression is significant [54].

It is not uncommon for *in vitro* concentrations to yield impressive changes in gene expression, but this is of no practical value if the compound exhibits poor bioavailability. Polyphenols commonly fall into this category, with bioavailability preventing the *in vivo* replication of *in vitro* outcomes when the same molecule is ingested in an oral dose form [70–72].

2.4. The History and Evolution of Sulforaphane Research. It is twenty-five years since the identification and isolation of the transcription factor, Nrf2 (coded by the gene *nuclear factor erythroid 2-related factor 2*), was first described in the scientific literature [73]. In the ensuing years, Nrf2 has become a focus of active research on mechanisms of defence in mammalian cells; Figure 2 illustrates the upward trend in SFN research over the period [74]. The role of Nrf2 in human cells is very relevant to the subject matter of this review because SFN significantly activates Nrf2 and as such has the potential to modulate the expression of genes associated with redox

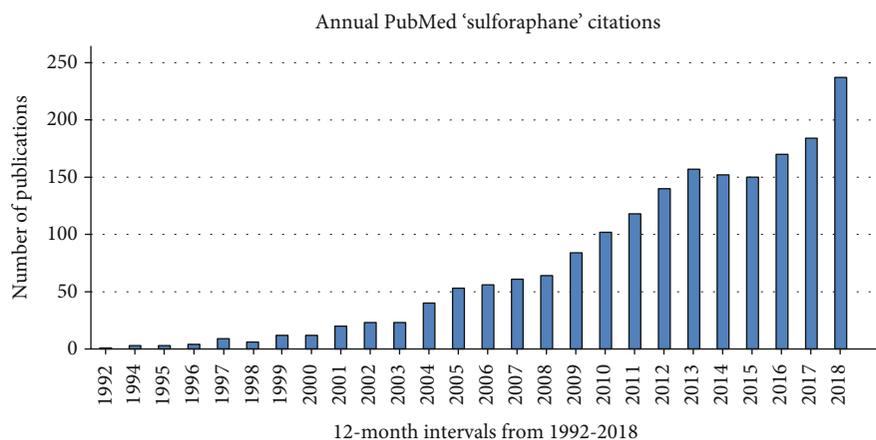


FIGURE 2: Sulforaphane research timeline; PubMed.

balance, inflammation, detoxification, and antimicrobial capacity, all key components of the *upstream* cellular defence processes [75].

There are many factors that can activate Nrf2. In addition to diet-derived molecules, a range of environmental stressors function as signals to activate Nrf2 and consequent expression of a battery of defensive genes [76]. Numerous commonly ingested phytochemicals are Nrf2 activators, and the reader is referred to a detailed discussion of the chemical properties and the subtle differences of individual phytochemical Nrf2 activators in relation to their interactions within relevant biochemical pathways in human cells [77].

In addition to providing a list of the more extensively studied phytochemical Nrf2 activators, Egger and Savinov suggest, in their concluding remarks, that although it is unlikely that a single phytochemical will emerge as a magic bullet for disease prevention or amelioration, future prospects could include phytochemical “cocktails” formulated for their synergistic effects [77]. In this regard, a larger quantity of low potency Nrf2-activating phytochemicals may provide the same effect as smaller quantities of a single Nrf2 activator such as SFN. If an additive or a synergistic effect of multiple Nrf2 activators provides significant Nrf2 activation, it may explain why diets rich in plant foods have been shown epidemiologically to significantly benefit human health [78].

Interest in SFN as a food-derived compound with significant clinical potential began in 1992 when a group [79] at Johns Hopkins University published its findings. The group had published two papers to support their research on the induction of anticarcinogenic enzymes derived from broccoli and on assay methods to rapidly detect such enzymes [79, 80]. Interestingly, SFN was identified here as a potent activator of cellular defence mechanisms approximately two years before the isolation of Nrf2 by Moi et al. [73] and Zhang et al. [79].

Broccoli-derived SFN was capable of activating the cytoplasmic transcription factor, Nrf2, which in turn translocated to the nucleus to activate the Antioxidant Response Element (ARE) in the promoter region of several hundred identified genes [58, 66, 81, 82]; many of which are related to cellular defence processes.

The Johns Hopkins group had found that the 3-day germinated broccoli seed contained 20-50 times more of the precursor glucoraphanin (GRN) than did the mature broccoli vegetable [81]. It was this finding that enabled the design of trials which could achieve clinically relevant SFN effects with small practical doses of dried broccoli sprouts.

3. Sulforaphane: Structure-Function Relationship

3.1. Physical Properties of Sulforaphane as an Intervention Compound. SFN is naturally derived from certain species of the *Brassica* vegetable family [83], most notably broccoli. Classified as cruciferous vegetables, they are known for their disease-preventive effects [84, 85]. When ingested, the bioactivity of crucifers is dependent on the dual presence of a precursor molecule, a *glucosinolate*, and an enzyme, *myrosinase*, which hydrolyses the precursor; the product is an isothiocyanate (Figure 3) [86].

Broccoli has been shown to be the most significant dietary source [87] of the precursor glucosinolate, GRN, which, in the presence of the *myrosinase* enzyme, is metabolised to SFN. Young sprouted broccoli seeds in the order of 3-7 days' growth have been shown to contain the highest GRN levels [81].

The structure of this small molecule (M.W. 177.29 and $\log P = 0.23$) confers upon SFN some unique advantages not afforded other phytochemicals such as the polyphenols which are structurally large and essentially hydrophilic [70]. One of the major advantages for SFN is its higher bioavailability, a consequence of its structure and lipophilicity (Figure 4).

3.2. Bioavailability: Relationship to Molecular Structure. SFN has been demonstrated to have an absolute bioavailability of around 80% [57] and to peak in the bloodstream around 1 hour following ingestion [88, 89]. By comparison, the polyphenols which are large bulky higher molecular weight molecular structures typically exhibit bioavailability of around 1-8% [90].

For a food-derived molecule to achieve sufficient intracellular concentration to affect gene expression, its

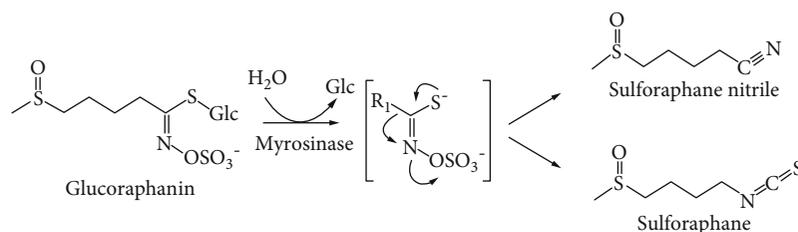


FIGURE 3: The synthesis of isothiocyanates via a hydrolysis reaction of the glucosinolate by the myrosinase enzyme. Sulforaphane is the isothiocyanate synthesised from the glucosinolate, glucoraphanin (an image adapted from Dinkova-Kostova et al. [86]).

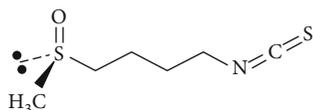


FIGURE 4: Sulforaphane ($C_6H_{11}NOS_2$)—molecular structure of sulforaphane (4-methylsulfinylbutyl isothiocyanate).

bioavailability must be high enough that it can be absorbed through the intestinal epithelium and the several other membranes between the gut and the target cell. With its high bioavailability, SFN can therefore be considered as having at least one of the key properties necessary to be considered for development as a nutraceutical compound.

4. Sulforaphane as a Molecule with Nutrigenomic Properties

Nrf2 has been variously described by several researchers as an “activator of cellular defence mechanisms” [91], “the master redox switch” [92], and “a guardian of health span and gatekeeper of species longevity” [93]. As a mediator for amplification of the mammalian defence system against various stressors, Nrf2 sits at the interface between our prior understanding of oxidative stress and the endogenous mechanisms cells use to deal with it [54].

What is emerging is that diseases known to be underpinned by oxidative stress are proving to be more responsive to such amplification of cellular defences via Nrf2 activation than by administration of direct-acting antioxidant supplements [22, 94].

4.1. The SFN-Nrf2: Activating Gene Expression in Cellular Defences. The essence of a very complex biochemical process [74] is that in its basal state, Nrf2 is sequestered to Kelch-like ECH-associated protein 1 (Keap-1) and associated with cytosolic actin filaments; however, when Keap-1 detects a stressor which may threaten the cell’s integrity, activation of the complex leads to a dissociation of Nrf2 from Keap-1 [95]. Hereafter, it translocates to the nucleus where it may induce expression of its many target genes, aligning with the ARE in the promoter region of these genes. The ARE is a *cis*-acting enhancer sequence that is upstream of many Phase 2 detoxification and antioxidant genes [96] (Figure 5 [97]).

Loss of the Nrf2-ARE function in mice has been shown to increase susceptibility [98] to acute toxicity, inflammation,

and carcinogenesis due to the inability to mount adaptive responses. The elucidation of this process showed that the activation of Nrf2-ARE induces a large battery of cytoprotective enzymes [99].

Cellular Nrf2 levels are under strict control by multiple mechanisms but the best-characterised is the one which is mediated by interaction with Keap-1 [63]. Keap-1 not only binds Nrf2 to cytoplasmic actin filaments in the basal state but it also acts as a sensor, especially of subtle redox changes in the cell.

The chemistry of sulfur plays an integral role in Nrf2 activation and subsequent modulation of gene expression. All Nrf2 activators react with thiol groups. Keap-1 is rich in sulfur-rich cysteine residues [99] and is under oxidation-reduction (and alkylation) control via its highly reactive thiol groups.

An inducer such as SFN activates the Nrf2-Keap-1 complex, with sulfur chemistry playing an important role [99].

4.2. The Significance of the Nrf2-SFN Relationship. Nrf2 is ubiquitously expressed with the highest concentrations (in descending order) in the kidney, muscle, lung, heart, liver, and brain [73, 100]. The activation of Nrf2 activators has been found to be closely associated with their molecular structure [100–102]. Because food-derived SFN is readily bioavailable, such universal Nrf2 tissue distribution enhances SFN’s potential to modulate systemic gene expression [92].

The properties of Nrf2 are such that it can be considered a novel drug target with potential applications across a broad range of conditions. Interestingly, the Nrf2-activating properties of SFN have been experimentally used in conjunction with pharmaceuticals. By way of an example, SFN’s effect on Nrf2 has been investigated in this context as a means of minimising the nephrotoxicity which typically limits the use of the chemotherapeutic drug, cisplatin [103]. Another example illustrates a synergistic antioxidant and anti-inflammatory response when SFN is combined with Exemestane, a synthetic steroidal inhibitor of the aromatase reaction that catalyses the terminal and rate-limiting step of the biosynthesis of estrogens. The combination may be considered to be protective against other chronic diseases unrelated to aromatase inhibition and the significance of such coadministration is expanded in Section 9.4 [104].

4.3. Pleiotropic Effects of SFN. Although SFN is most often considered for its Nrf2-dependent effects and largely

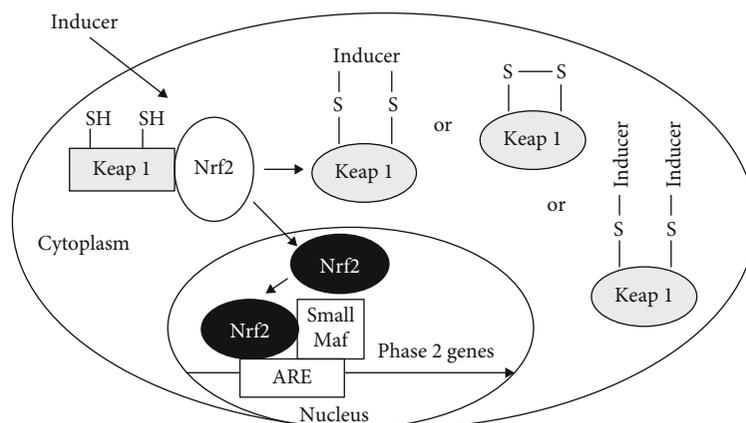


FIGURE 5: Mechanism by which an inducer affects expression of Phase 2 detoxification genes (an image from Zhang et al. [97]).

associated with the induction of antioxidant and Phase 2 detoxification enzymes, other less well-characterised mechanisms are associated with this phytochemical molecule. These Nrf2-independent mechanisms include but are not limited to the induction of apoptotic pathways, suppression of cell cycle progression, inhibition of angiogenesis and anti-inflammatory activity, and inhibition of metastasis, primarily relevant to cancer [62].

One such effect is its action as a histone deacetylase (HDAC) inhibitor [105, 106], and there is a growing focus on the role of SFN and other phytochemicals on such epigenetic effects [107, 108] and more recently on the role of SFN as an inhibitor of microRNAs [109]. Epigenetic effects are of particular clinical interest in that such changes are potentially reversible and thereby may provide an opportunity for intervention in earlier stages of the cancer process [110]. Tumour suppressor genes such as p53 may be epigenetically inhibited [111] so that therapies aimed at removing such suppression are attractive options, especially if they can be available through dietary means.

No discussion of SFN and Nrf2 would be complete without reference to the fact that both Nrf2 activators and Nrf2 inhibitors can be utilised in cancer therapy. A very recent paper [112] highlights this dual role and its implications for Nrf2 activation. It suggests that because Nrf2 can modulate the detoxification pathways, its effect on anticancer drugs may lead to chemoresistance and that the switch between a beneficial and a detrimental role for Nrf2 in cancer cells depends on a number of factors which include the tight control of its activity. This poses an obvious dilemma which is already under active discussion and investigation [113–115]; SFN and other phytochemicals capable of modulating Nrf2 form part of such investigation [112].

A 2012 gene expression study to evaluate the effect of SFN as an Nrf2 activator showed that despite the very large 5- to 20-fold increase in Nrf2 binding at their AREs, only a small increase in expression signal was observed. The researchers concluded that there may be other determinants, such as tissue-specific cofactors, negative feedback loops, and epigenetic or signalling mechanisms, which affect both basal expression and Nrf2-mediated transcriptional regulation of these highly expressed genes in cells [116].

4.4. Major Actions of SFN at the Cellular Level. The major documented cellular actions of SFN are listed in the non-exhaustive summary shown in Table 2 along with commentary on their clinical implications. These *upstream* processes have significant *downstream* effects and are associated with the observed effects in clinical trials using SFN or a dietary source of SFN. Most but not all of these actions are associated with Nrf2 activation.

5. Sulforaphane in Core Cellular Processes

5.1. Multiple Gene Targets and the Nrf2/ARE Pathway. It has been suggested that well in excess of 500 genes have been identified as being activated by SFN via the Nrf2/ARE pathway [132–134], and it is likely that this underestimates the number as others are being discovered.

The large battery of upregulated cytoprotective genes includes those coding for the endogenous enzyme and non-enzyme antioxidants as well as Phase 2 detoxification enzymes [58]. Nrf2 plays a crucial role in the coordinated induction of those genes encoding many stress-responsive and cytoprotective enzymes and related proteins [135]. These include NAD(P)H:quinone reductase-1 (*NQO1*), haemoxygenase-1 (*HO-1*), glutamate-cysteine ligase (*GCL*), glutathione-S-transferase (*GST*), glutathione peroxidase (*GPX1*), thioredoxin (*TXN*), thioredoxin reductase (*TXNRD1*) [92], and PPAR- γ (*PPARG*) [136].

These endogenously-generated enzyme and nonenzyme molecules are not generally considered to necessarily function as “antioxidants” even though they exhibit significant redox-modulating capacity *as* and *when* the cell requires it.

When Zhang and colleagues [79] of the Johns Hopkins group were investigating chemoprevention in the early 1990s, they had been working on cytoprotective genes including those coding for the Phase 2 detoxification enzymes *NQO1* and the *GST* families; the discovery that these genes were significantly induced by broccoli sprout-derived SFN provided the foundation for the rapid interest in research in this field.

Of the available SFN clinical trials associated with genes induced via Nrf2 activation, many demonstrate a linear dose-response (Table 3). More recently, it has become

TABLE 2: Summary of clinically relevant actions of SFN.

Action	Clinical implications
(1) Increases synthesis of glutathione [117].	This has implications for oxidative stress and detoxification as glutathione is the substrate for both pathways. Glutathione is also an antioxidant in its own right.
(2) Inhibits some Phase 1 detoxification enzymes that activate chemical carcinogens [118].	This reduces the level of toxic intermediates with carcinogenic potential. It also allows Phase 2 to “keep pace” with Phase 1 processing.
(3) Increases activity of Phase 2 detoxification enzymes. Sulforaphane is considered the most potent of the Phase 2 inducing substances [79].	As a monofunctional inducer, sulforaphane is considered to be a significant component of the anticarcinogenic action of broccoli.
(4) Provides significant antioxidant activity, largely due to its ability to induce glutathione synthesis.	Glutathione is a critical factor in protecting organisms against toxicity and disease [119]. The ability of sulforaphane to upregulate glutathione synthesis is highly significant.
(5) Acts as a histone deacetylase inhibitor, providing DNA protection [120–122].	Development of histone deacetylase inhibitors is a key avenue for cancer drug research.
(6) Induces apoptosis, inhibits MMP-2 (metastasis), and inhibits angiogenesis and cell cycle arrest [28, 105, 123, 124] (interacts at several levels).	Therapeutic interventions which exhibit several related actions targeting the same underlying defect are considered highly desirable.
(7) Limits proinflammatory effects of diesel chemicals by upregulation of Phase 2 enzymes [125].	Environmental pollutants are known to contribute to various lung diseases. Removal of the toxins reduces tendency to disease.
(8) Induces thioredoxin (Trx) as part of the ARE.	Thioredoxin is implicated in cardioprotection by triggering several <i>survival</i> proteins [126]. Sulforaphane may have beneficial effects in cardiovascular disease.
(9) Bactericidal against <i>Helicobacter pylori</i> and also blocks gastric tumour formation in animals [127].	<i>Helicobacter</i> is known to contribute to development of stomach cancer. Elimination of the organism without the use of typical antimicrobial <i>Triple Therapy</i> could protect the colonic microflora.
(10) Protects dopaminergic cells from cytotoxicity and subsequent neuronal death (cell culture) [128].	Dopaminergic neurones are associated with Parkinson’s disease. Pharmaceuticals to treat Parkinsonism are not without risk and the disease is not usually detected until more than 50% of the neurones have been lost. A chemoprotective tool could prevent premature loss.
(11) Increases p-53 (associated with tumour suppression) and bax protein expression, thereby enhancing cellular protection against cancer [129].	Sulforaphane is an attractive chemotherapeutic agent for tumours with a p53 mutation [62].
(12) Limits effect of aflatoxin on liver cells [26].	Interventions which can offer significant protection against environmental and food-borne pollutants could prevent the consequences of these factors. Appropriate doses of sulforaphane-yielding substances are yet to be determined.
(13) Enhances natural killer cell activity and other markers of enhanced immune function [117].	The immune system is a critical part of the body’s defences against inflammatory as well as infectious diseases. Most diseases benefit from enhancement to immune function.
(14) Suppresses NF- κ B, a key regulator of inflammation [117]. NF- κ B expression is downregulated by sulforaphane and as such downregulates inducible proinflammatory enzymes such as cyclooxygenase (COX-2) and NO synthase (iNOS).	As an inhibitor of NF- κ B as well as an activator of Nrf2, SF modulates many cancer-related events, including susceptibility to carcinogens, cell death, cell cycle, angiogenesis, invasion, and metastasis [117].
(15) Sulforaphane is not directly antioxidant. Instead, it exhibits a weak <i>prooxidant</i> effect [130].	Because sulforaphane is not directly antioxidant but exerts its antioxidant effect primarily by induction of glutathione and other antioxidant compounds, it is considered to exhibit an <i>indirect</i> antioxidant effect.
(16) Potent inducer of HO-1 (haemoxygenase-1).	Haemoxygenase-1 plays an important role in modulating the effects of oxidants in the lungs [131].

apparent that SFN can behave hormetically [137] with different effects responsive to different doses. This is in addition to its varying effects on different cell types and consequent to widely varying intracellular concentrations [125, 138–142].

5.2. SFN as a Redox Modulator. Even though enzymes known to function within the Phase 2 detoxification pathway are not typically considered to be “antioxidants,” it has now been firmly established that NQO1 provides major antioxidant functions by virtue of its obligatory two-electron reduction

TABLE 3: Sulforaphane dosage from lowest to highest in selected clinical trials.

Condition	~Daily SFN dose	First author	Year
Equivalent sulforaphane dose			
Autism	9-14 mg (50.8–79.0 μmol)	Singh et al. [228]	2014
Nasal allergic response	18 mg (101.5 μmol)	Heber et al. [230]	2014
Asthma	18 mg (101.5 μmol)	Brown et al. [226]	2015
Chronic obstructive lung disease	19 mg (107.2 μmol)	Riedl et al. [29]	2009
<i>Helicobacter pylori</i> infection	30 mg (169.2 μmol)	Yanaka et al. [217]	2009
Gastric mucosal repair	30 mg (169.2 μmol)	Yanaka A. [216]	2011
Detoxification (atmospheric pollution)	36 mg (203.0 μmol)	Egner et al. [231]	2011
Type 2 diabetes	40 mg (225.6 μmol)	Bahadoran et al. [232, 233]	2012
Prostate-specific antigen (PSA) doubling time	60 mg (338.4 μmol)	Cipolla et al. [229]	2015
FRESH BROCCOLI SPROUTS			
<i>Helicobacter pylori</i> infection	14-56 grams of fresh sprouts	Galan et al. [218]	2006
Inflammation markers in overweight	30 grams of fresh sprouts	Lopez-Chillon [209]	2018
Metabolic syndrome	100 grams of fresh sprouts	Murashima et al. [25]	2004
Glucoraphanin as myrosinase-inactive broccoli “extract”			
No prevention with 6 pills branded “extract”	180 mg (0.41 mmol) GRN—not SFN	Atwell et al. [234]	2015
Sulforaphane supplement—a null response trial			
<i>Helicobacter pylori</i> infection	2 mg (11.28 μmol)	Chang et al. [235]	2015

mechanism which diverts quinones from participating in oxidative cycling and generation of reactive oxygen intermediates.

A major new perspective on the functional importance of this enzyme [143–145] followed the finding that the gene coding for NQO1 is highly inducible and that its increased induction protected animals and their cells against oxidative stress [143–145]. SFN is considered to be one of most potent phytochemical inducers of NQO1 [96, 146]. As such, SFN’s nutrigenomic effects contribute to the enhancement of the cell’s antioxidant capacity [64]. Figure 6 illustrates the comparative induction of SFN and other phytochemicals.

5.3. Endogenously Generated “Antioxidants” in Type 2 Diabetes. Given the role of SFN in induction of Nrf2-dependent cytoprotective genes, SFN might be a useful candidate for modulation of *upstream* genes associated with the etiology of T2DM. A 2016 review paper reaffirms a rationale for the “unifying hypothesis” proposed by Brownlee in 2001 in which generation of ROS is the key central theme linking the pathogenesis of T2DM and CVD [147]. In further support of this hypothesis, Rask-Madsen and King reinforce the possibility that endogenous protective pathways could protect against vascular complications in T2DM [148]. The following sections highlight the role of several inducible redox-modulating molecules with reference to their activity in T2DM.

5.4. Highlighting Redox-Modulating Nrf2 Target Genes. Several well-studied Nrf2-dependent target genes of possible relevance are those encoding synthesis of glutathione (GSH), Trx, HO-1, and NQO-1. Each has been shown to be induced by SFN in a variety of cell types, including endothelial cells. A study [149] using human aortic cells showed that the activa-

tion of the Nrf2-ARE pathway may represent a novel therapeutic approach for the treatment of inflammatory diseases such as atherosclerosis.

In support of this approach, a 2009 combined cell culture/animal study [150] showed that shear stress in blood vessels keeps Nrf2 in an activated state and as such protects against endothelial dysfunction. Activated by SFN, Nrf2 was shown to prevent endothelial cells from exhibiting a pro-inflammatory state via the suppression of p38-VCAM-1 signalling, providing a novel therapeutic strategy to prevent or reduce atherosclerosis.

In other tissues of the cardiovascular system, Nrf2 has been shown to regulate both basal and inducible ARE-controlled cytoprotective genes in cardiomyocytes [151]. As with endothelia, Nrf2 is required for protection against glucose-induced oxidative stress and cardiomyopathy in the heart.

6. SFN: Its Redox-Modulating Effects

6.1. Glutathione. The nonenzyme antioxidant GSH is a major contributor to cellular redox status and the rate-limiting enzyme for its synthesis; glutamate-cysteine ligase (coded by the gene *GCL*) can be induced by SFN [152]. Antioxidants in general and glutathione in particular can be depleted rapidly under conditions of oxidative stress, and this can signal inflammatory pathways associated with NF- κ B [153]. Nrf2 has been found to be the primary factor inducing the cell survival system under GSH depletion [154]. Also of interest is the finding that Nrf2 transcriptional activity declines with age [155, 156], leading to age-related GSH loss among other losses associated with Nrf2-activated genes. This effect has implications too for decline in vascular function with age [157].

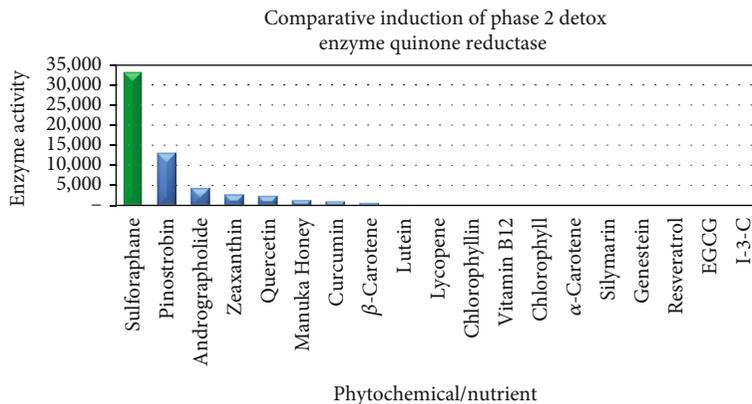


FIGURE 6: Comparison of capacity to induce NQO1 by a range of phytochemicals, indicating that SFN exhibits many-fold greater inducer ability (data compiled from Yang and Liu [168] and Fahey and Kensler, 2008).

Some of the age-related decline in function can be restored with Nrf2 activation by SFN [158]. Studies in aged mice showed that age-related changes in Th1 immunity could be restored using SFN as an intervention. This finding is compatible with the growing recognition of the importance of the Nrf2 pathway in innate immunity and has implications for human health [159]. A 2017 clinical pilot study examined the effect of an oral dose of 100 μ mol (17.3 mg) encapsulated SFN on GSH induction in humans over 7 days [158]. Pre- and postmeasurement of GSH in blood cells that included T cells, B cells, and NK cells showed an increase of 32%. Interestingly, the researchers found that in the pilot group of nine participants, age, sex, and race did not influence the outcome.

Disturbances of thiol-related mechanisms have been observed [160] in diabetes, with plasma levels of protein-bound thiols lower in T2DM than in controls. These thiols include GSH and Trx. An animal study [161] illustrates the relationship between depressed GSH and the development of atherosclerosis. In this experiment [7], the rate-limiting enzyme in GSH synthesis, *gamma-glutamyl-cysteine synthetase* (γ -GCS), was shown to be downregulated *early* in the atherosclerosis process. This effect preceded the appearance of lipid peroxidation products by several months. The antioxidant enzyme, glutathione peroxidase (GPx) was simultaneously downregulated.

Erythrocyte levels of GSH have been shown to change depending on the stage of the diabetic process of the individual [162]. It has been shown that compared to controls, pre-diabetic patients exhibit a significant lowering of GSH [163]. As the disease progresses to diabetes and later to diabetes with cardiovascular complications, GSH levels rise; however, they do not reach the levels of controls. The variability in GSH levels depending on the stage of the disease makes it difficult to use GSH as an effective clinical trial biomarker to measure change.

An infusion of GSH as an intervention in a clinical trial [164] was shown to reverse endothelial dysfunction by strongly potentiating the effect of acetylcholine-mediated vasodilation via enhanced nitric oxide activity. Because GSH as a tripeptide molecule is degraded by gastric proteo-

lytic enzymes, it is generally considered as being unsuitable as an oral therapeutic [165]. If SFN can be shown to induce GSH in endothelial cells, this may provide an alternative means of enhancing GSH levels in endothelial and pancreatic beta-cells with a view to reducing the complications of T2DM together with the many conditions for which dysregulated GSH is associated.

6.2. Thioredoxin: Protection from Elevated Blood Glucose. Thioredoxin (Trx) is a potent protein disulfide that participates in many thiol-dependent cellular reductive processes and plays an important role in antioxidant defence, signal transduction, and regulation of cell growth and proliferation. As a cellular thiol, Trx has been shown [166] to be associated with the development of diabetic complications. Like GSH, Trx has been shown to protect cells against high ambient glucose [167].

The thioredoxin system (Figure 7) consists of thioredoxin, thioredoxin reductase, and NAD(P)H.

Like GSH, Trx contributes to the cellular thiol pool [170] with the thioredoxin system shown to exhibit cardioprotective effects [171]. The pentose phosphate pathway can alleviate much of the oxidative stress created by excess glucose [169].

There are few studies to associate SFN with heart disease but significant cardioprotection was demonstrated in an animal study [126] using fresh broccoli homogenate. Changes included improved postischaemic ventricular function, reduced myocardial infarct size, and decreased cardiomyocyte apoptosis after the rats were sacrificed. These findings correlated with increased levels of Trx as well as HO-1.

A 1997 study [172] investigating the role of thioredoxin in vascular biology describes the induction of mitochondrial antioxidant enzyme, superoxide dismutase (MnSOD) by Trx. In addition, Trx influences hormones such as insulin as well as glucocorticoid receptors and other proteins such as endothelial nitric oxide synthase and signalling proteins such as transcription factors. The findings of a Phase 1 clinical trial [25] demonstrated that 100 grams of fresh broccoli sprouts over a 7-day period provided cardiovascular benefits which

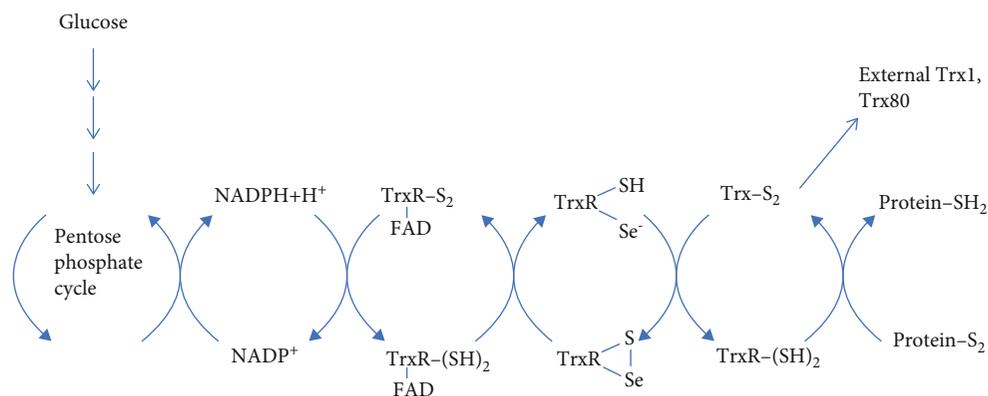


FIGURE 7: The thioredoxin system and its relationship with glucose metabolism in the pentose phosphate cycle. The pentose phosphate cycle generates reducing equivalents which are transferred along a series of cycling redox reactions. Induction of Trx and Trx reductase by SFN enables glucose to be metabolised as an alternative to the synthesis of superoxide radical to alleviate much of the metabolic stress associated with T2DM (a figure adapted from Holmgren and Lu [169]).

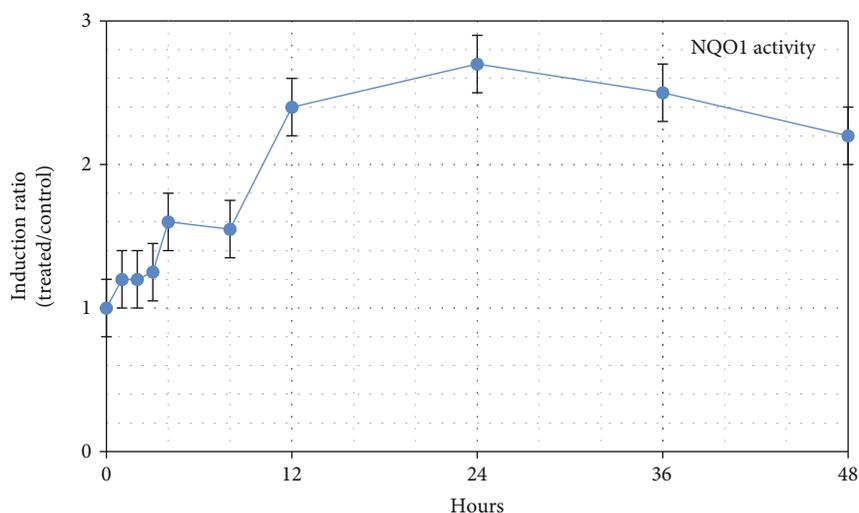


FIGURE 8: NAD(P)H quinone dehydrogenase 1 activity over time following sulforaphane ingestion (an image from Cornblatt [88]).

included favourable changes in blood lipids as well as reduction in biomarkers of oxidative stress. This study however did not assay the broccoli sprouts for their SFN yield, limiting its usefulness.

6.3. NAD(P)H Quinone Dehydrogenase 1: Beyond Redox Modulation. NAD(P)H quinone dehydrogenase 1 (coded by the gene *NQO1* and with the enzyme sometimes abbreviated as NQO1) is emerging as an Nrf2-target enzyme with broad cytoprotective properties. A paper [173] published almost two decades ago claims that *an extensive body of evidence supports the conclusion that catalysing obligatory two-electron reductions of quinones to hydroquinones, NQO1, protects cells against the deleterious effects of redox cycling of quinones and their ability to deplete glutathione.* The same researchers [144] have since published on this topic discussing what they describe as a *“multifunctional antioxidant enzyme and exceptionally versatile cytoprotector.”* They suggest too that NQO1 with cytoprotective roles which extend well beyond its catalytic function could be considered as a

“marker cytoprotective enzyme.” Further, they state that *NQO1 is one of the most consistently and robustly inducible genes among members of the cytoprotective proteins.*

6.4. NQO1 Pharmacokinetics following SFN Ingestion. A study used breast tissue to measure the pharmacokinetics of NQO1 induction over 24 hours, following a single serve of a broccoli sprout homogenate (SFN = 200 μ mol) one hour prior to mastectomy [88].

Maximal induction of NQO1 occurred at around 24 hours, declining thereafter (Figure 8). This peak represents an approximate 2.8-fold induction over baseline. These findings are useful when considering the effect of SFN as an intervention material in acute compared with chronic conditions. A significant increase in NQO1 occurred between 6 and 12 hours, a timeframe that may not be sufficiently responsive for management of an acute state, leaving one to conclude that NQO1 induction is best suited to chronic conditions where a rapid response may not be necessary.

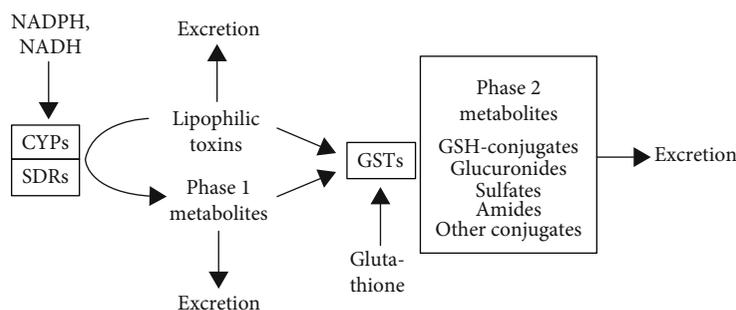


FIGURE 9: Interaction of Phase 1 and Phase 2 metabolites in detoxification (an image from McElwee et al. [192]).

6.5. Comparative Phytochemical NQO1 Induction. The induction of NQO1 has been investigated in different studies to compare the effect of well-known phytochemicals [146, 168, 174]. The comparatively much higher NQO1 induction by SFN against popular plant-derived supplements is evident [146].

It has been claimed here and elsewhere that SFN is the most potent naturally occurring inducer [146, 175] of this enzyme (Figure 6) NQO1's antioxidant capacity extends to scavenging superoxide directly [176], albeit not as efficiently as does SOD.

6.6. NQO1: Recycling Cellular Bioactives. NQO1's other functions extend to the maintenance of coenzyme Q 10 and vitamin E in their active reduced forms [144]. Induction of NQO1 by SFN also coordinately induces [58] genes encoding cellular NADPH-regenerating enzymes such as glucose-6-phosphate dehydrogenase, 6-phosphogluconate dehydrogenase, and malic enzyme. NADPH in turn assists in maintaining GSH in its reduced state. The NQO1 enzyme provides major antioxidant functions by virtue of its two-electron reduction mechanism; this diverts quinones from participating in oxidative recycling and production of ROS and prevents mutagenic changes to DNA [144, 177, 178]. This function is clinically relevant to chemoprevention.

6.7. Haemoxygenase-1 (HO-1). HO-1 is an inducible isoform of the first and rate-controlling enzyme of the degradation of haem into iron, carbon monoxide, and biliverdin, the latter being subsequently converted into bilirubin [179]. HO-1 is considered to have potent cytoprotective effects which include antioxidant and anti-inflammatory properties in cardiovascular and other tissues. It has been suggested that cytoprotection may be due to bilirubin directly inhibiting NADPH oxidase activity, thereby reducing superoxide generation [180].

Although the mechanism for the anti-inflammatory effect of HO-1 has not been fully elucidated, there are known associations between HO-1 and a number of cytokines. The 5'-flanking region of the *HO-1* gene contains binding sites for the transcription factors that regulate inflammation, including NF- κ B and activator protein-1 (AP1) [181]. Leukocyte *HO-1* gene expression is significantly lower in patients with and without diabetic microangiopathy compared with control subjects and normalization of blood glucose results

in a reduction in HO-1 antigen in the cytoplasm of mononuclear leukocytes [182].

Hyperglycaemia is known to increase the formation of advanced glycation end products (AGEs). In endothelial cells, the interaction of the AGE with its receptor, RAGE, induces generation of ROS, NF- κ B translocation, and expression of several proinflammatory and procoagulatory molecules [183]. In normal cells, RAGE is present at low levels but is increased in the endothelia of diabetics [180].

Given the theme of the above discussion, it could be asked whether the redox-inflammation couple could be the common upstream factor at play in a number of chronic diseases, of which T2DM is an example. It has, in fact already been proposed [7, 184] that oxidative stress is the pathogenic mechanism linking insulin resistance with dysfunction of both pancreatic beta-cells and the endothelium, eventually leading to overt diabetes and cardiovascular disease.

6.8. Redox Effects in Phase 1 vs. Phase 2 Detoxification Pathways. As long ago as 1985, it was determined that the ideal chemoprotective compounds are monofunctional inducers of Phase 2 detoxification enzymes. Monofunctional inducers function by metabolising the oxidative and carcinogen-activating products of the Phase 1 enzymes, without having any significant effect on Phase 1 activity itself [185]. Toxins presented to the Phase 1 enzymes produce intermediate compounds which are sometimes more toxic to cells than the initial toxin. It is therefore important that Phase 2 is sufficiently active that the intermediate products cannot accumulate in the cellular environment. The majority of chemical carcinogens require metabolic activation by Phase 1 before they can initiate cancer [186]. Figure 9 illustrates the Phase 1 and Phase 2 detoxification pathways [187].

As a monofunctional inducer, SFN has been described an ideal detoxifier, as its effect on Phase 1 is minimal compared with its significant activity on Phase 2 [188]. By comparison, many of the most potent of the synthetic SFN analogues [189] are bifunctional inducers and not the monofunctional inducers having the most chemopreventive effect. Several synthetic compounds [190] have been investigated for their chemopreventive potential against lung cancer in smokers [191].

The process of cellular detoxification of both exogenous and endogenous factors entails two phases: Phase 1 (oxidative activation reactions) and Phase 2 (conjugative reactions), effected by several large and diverse gene families [192].

6.9. Significance of Induction of Phase 1 and Phase 2 Detoxification Enzymes. Not all Brassica-derived compounds are monofunctional inducers. Indole-3-carbinol (I-3-C) derived from the mature broccoli vegetable is a bifunctional inducer and as such may lead to the generation of highly toxic intermediate compounds which may overwhelm the capacity of the localised direct-acting antioxidants to quench them or the Phase 2 processes to detoxify them [193].

By contrast, SFN selectively upregulates Phase 2 detoxification enzymes, minimising the risk of generating excessive amounts of reactive intermediates (Figure 9 [192]). As a consequence, although some I-3-C animal studies show an anticarcinogenic effect, other studies using I-3-C show it to have carcinogenic potential where comparable studies using SFN do not [194–196]. It should be noted that the comparatively small quantity of I-3-C generated from the glucosinolates in broccoli vegetable is unlikely to replicate the effects of isolated synthetic I-3-C concentrations used in cell culture studies [87].

7. SFN: Its Anti-Inflammatory Effects

7.1. Regulation of NF- κ B. Members of the NF- κ B family of transcription factors function as dominant regulators of inducible gene expression in virtually all cell types in response to a broad range of stimuli, with particularly important roles in coordinating both innate and adaptive immunities [197], as well as inflammatory responses, cell differentiation, proliferation, and apoptosis.

NF- κ B is controlled by various mechanisms of posttranslational modification and subcellular compartmentalisation as well as by interactions with other cofactors or corepressors [198]. The NF- κ B family of transcription factors includes RelA (p65), RelB, and others and as a complex, NF- κ B mediates immune responses to cellular challenges that include bacterial and viral infection and inflammation [63].

The activity of NF- κ B is tightly regulated at multiple levels, a factor that may be associated with its influence on the expression of numerous genes [199]. Nuclear translocation of NF- κ B is primarily controlled by signalling associated with I κ B kinase (IKK) in two related pathways associated respectively with the NF- κ B classical (canonical) and alternative pathways.

Among the most potent NF- κ B activators are tumour necrosis factor (TNF- α), interleukin (IL)-1 β and bacterial lipopolysaccharide (LPS), with TNF- α activation being one of the best characterised of the NF- κ B signalling pathways [200].

7.2. The Action of NF- κ B in Intestinal Epithelial Cells. Nrf2 and NF- κ B are both well-studied cellular transcription factors, and their effects occur in all cells including those of the intestinal epithelium. The gut-immune interface describes the signalling network that connects the intestinal epithelial cells to the immune cells of the lamina propria, situated directly below the epithelium [201]. Here, the microbiota, via this interface, also influence immune function including inflammatory pathways. As such, the gut-immune interface directly connects the cellular functions of

redox-balance, inflammation, and infection control via immune modulation.

7.3. SFN at the Gut-Immune Interface. Because SFN has been shown to inhibit NF- κ B in endothelial cells [202], it is likely the same effect would occur in other epithelial cells such as the intestinal epithelium, thereby retarding local inflammation.

Whereas SFN directly activates cytosolic Nrf2, its action on NF- κ B is to inhibit NF- κ B binding to the DNA [203]. NF- κ B plays a key role in the immune system where it is activated by a series of events initiated by Toll-like receptors (TLR) on epithelial cells [204]. TLR2 and TLR4 can identify distinct molecular patterns on the cell wall of invading pathogens. These patterns act as innate sensors but also shape and bridge innate and adaptive immune responses.

7.4. Cross Talk between Nrf2 and NF- κ B. SFN is associated with cellular defences via mechanisms governed by the transcription factors Nrf2 and NF- κ B; molecular cross talk between these transcription factors has been reported [63]. Imbalance between Nrf2 and NF- κ B is associated with a significant number of diseases across various body systems, and these relationships are the subject of extensive research in cancer biology in particular [205].

Although the complex interplay between Nrf2 and NF- κ B has been highlighted, there remains much to be explored in order to understand how such relationships may impact disease pathophysiology at the molecular level. As part of the cross talk between these two transcription factors, NF- κ B has been shown to regulate Nrf2-mediated ARE expression. Several mechanisms exist by which p65 (the canonical NF- κ B subunit) can exert negative effects on ARE-linked gene expression [206]. It would seem that the cross talk between Nrf2 and NF- κ B enables cells to more finely regulate their responses to cellular stressors.

7.5. Immune Modulation (Anti-Inflammatory Effects). Activation of TLR4 by the endotoxin released by gram-negative bacteria results in signalling that activates NF- κ B with subsequent generation of inflammatory cytokines [204]. Toll-like receptor (TLR4) pathways mediate proinflammatory cytokine and interferon responses [207]. SFN has been shown in a thiol-dependent manner to suppress TLR4 oligomerization. Saturated fatty acids are known to act as ligands for TLR4 in macrophages and adipocytes, with these signals in turn regulating various proinflammatory transcription factors [208]. More recently, in-depth investigation of the microbiome has uncovered the pathways that link these very signals to cardiometabolic effects, thereby connecting the gut-immune relationship to systemic disease [13].

7.6. Effect of SFN on Inflammation Markers in Humans. In a recent study using 30 grams of fresh broccoli sprouts incorporated daily into the diet, two key inflammatory cytokines were measured at four time points in forty healthy overweight people [209]. The levels of both interleukin-6 (IL-6) and C-reactive protein (CRP) declined over the 70 days during which the sprouts were ingested. These biomarkers were measured again at day 90, wherein it was found that IL-6

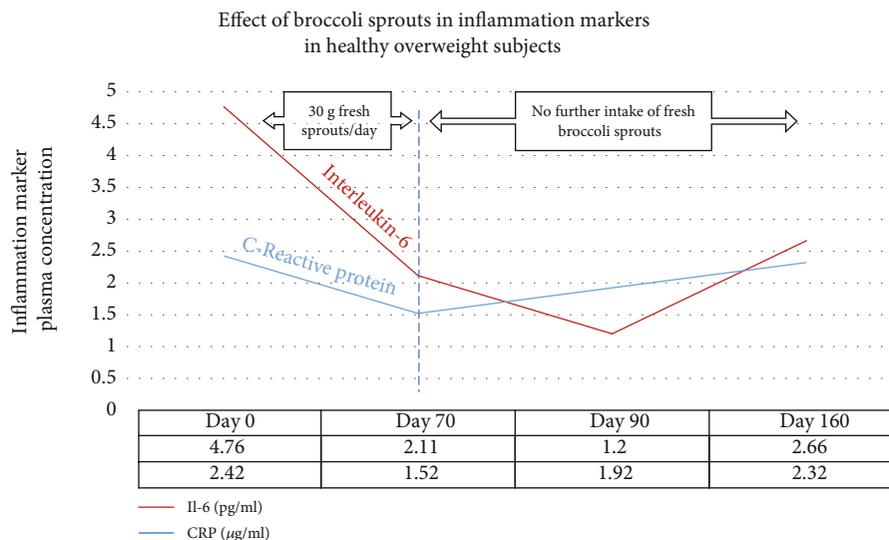


FIGURE 10: Effect of broccoli sprouts in inflammation markers in healthy overweight subjects (data from Lopez-Chillon et al. [209]).

continued to decline, whereas CRP climbed again. When the final measurement was taken at day 160, CRP, although climbing, had not returned to its baseline value. Il-6 remained significantly below the baseline level at day 160.

The sprouts contained approximately 51 mg (117 μ mol) GRN, and plasma and urinary SFN metabolites were measured to confirm that SFN had been produced when the sprouts were ingested. The data from this study are expressed visually in Figure 10.

7.7. Effect of SFN on Inflammation Markers in Type 2 Diabetes Patients. Where the study described above by Lopez-Chillon et al. investigated healthy overweight people to assess the effects of SFN-yielding broccoli sprout homogenate on biomarkers of inflammation, Mirmiran et al. in 2012 had used a SFN-yielding supplement in T2DM patients [210]. Although the data are not directly comparable, the latter study using the powdered supplement resulted in significant lowering of Il-6, hs-CRP, and TNF- α over just 4 weeks. It is not possible to further compare the two studies due to the vastly different time periods over which each was conducted.

8. SFN: Its Antimicrobial Effects

The complex signalling mechanisms discussed above will apply in a general sense to the modulation of core upstream processes that occur in human cells in general. In the following section, specific actions by SFN exhibit an antimicrobial effect on a common gut pathogen. It is not known at this stage whether the mechanisms are applicable to eradication of other pathogens with similar characteristics.

8.1. SFN and *Helicobacter pylori* Gut Infection. Although a direct antimicrobial effect has been demonstrated for extracts of cruciferous vegetables [211], the effect is not considered clinically relevant. More recently, SFN has been shown mechanistically and clinically to have a direct bac-

tericidal effect on the *Helicobacter pylori* bacterium via two separate mechanisms.

Approximately half of the global population is thought to be colonised by the *H. pylori* organism, making its classification as either a pathogen or a commensal uncertain; as such, it is sometimes described as a pathobiont [212]. *H. pylori* is shown to be symptomatic in some people and not in others, indicating that there may be individual control mechanisms that keep the organism in check. The popular dietary practice of salting food can also contribute to its pathogenicity. Sodium chloride, in the presence of *H. pylori*, becomes a cancer promoter, enhancing chronic gastric mucosal membrane inflammation [213].

H. pylori infection may be asymptomatic but by raising the pH of the gastric contents via continuous synthesis of ammonia, it contributes to impaired protein digestion and macromineral malabsorption. Iron absorption is well known to be impaired in the presence of *H. pylori* [214].

Consideration of the *upstream* processes that cells use to maintain homeostasis might indicate that the redox-inflammation couple might be associated. Recently, Yanaka, who had undertaken some of the earlier *H. pylori* trials using SFN as an intervention, reviewed several of the mechanisms by which Nrf2 activators may exhibit their antimicrobial effect [215]. Yanaka argues that significant protection to the gastrointestinal tract is afforded by the modulation of oxidative stress and inflammation as a result of simultaneous activation of Nrf2 and downregulation of NF- κ B [216].

In their 2009 study, Yanaka et al. demonstrated that broccoli sprouts suppressed the upregulation of the inflammatory markers, TNF- α and IL-1 β in the gastric mucosa by *H. pylori* infection in a wild type but not in Nrf2-/- mice, suggesting a systemic protective effect against gastritis that was the result of Nrf2 activation [217].

Over the past fifteen years, two clinical trials have demonstrated SFN's bactericidal effect on the *H. pylori* organism, a bacterium which is associated with gastric reflux and cancer [217, 218]. In the initial study, forty-eight *H. pylori*-infected

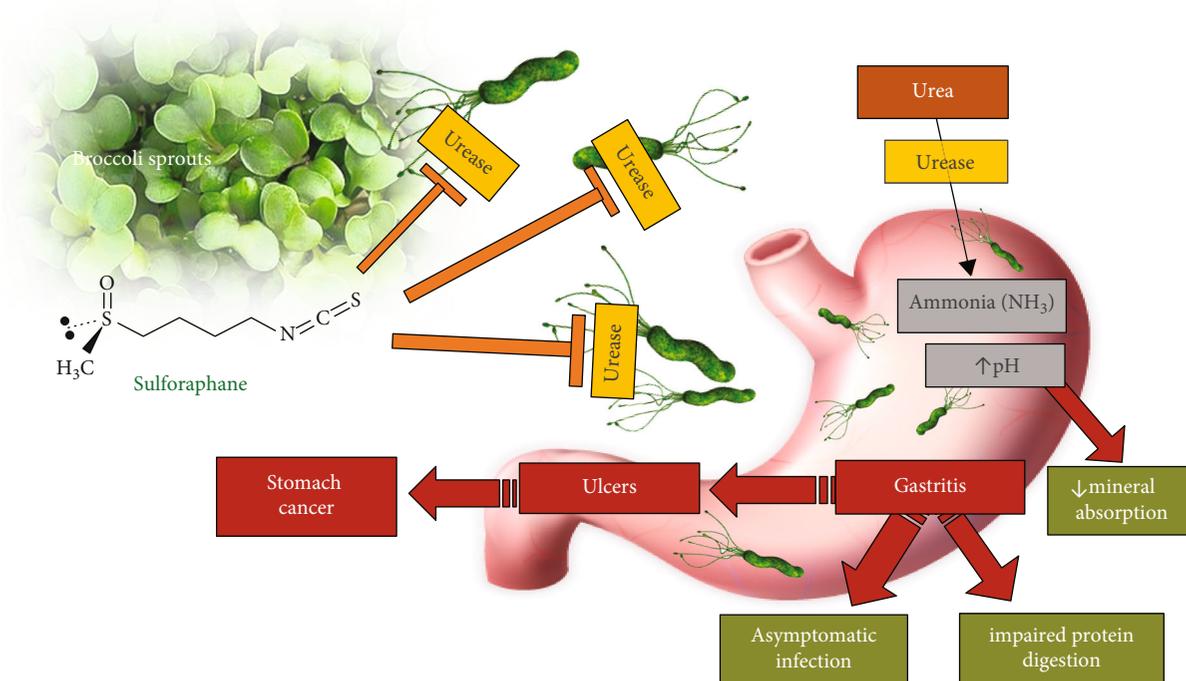


FIGURE 11: The proposed mechanism by which SFN inhibits urease synthesis by *H. pylori* and, in so doing, reduces the disease risks associated with *H. pylori* infection (an image adapted from Fahey et al. [220]).

subjects were given 70 grams fresh broccoli sprouts daily [217]. Three markers of *H. pylori* infection declined within eight weeks to below the diagnostic cutoff point. However, once the intervention had stopped, the levels of *H. pylori* returning to baseline levels after 8 weeks.

8.2. Urease Inhibition as a Mechanism for Regulating *H. pylori* Colonies. In extending the earlier SFN—*H. pylori* research, the urease-positive nature of the *H. pylori* gram-negative bacterium has been investigated. Urease activity in human and animal cells can be the cause of some pathogen-induced infections, and the ongoing quest to provide appropriate urease inhibitors includes the search for natural sources [219]. SFN has been demonstrated to exhibit urease activity, thereby potentially providing a clinical alternative to pharmaceutical antibiotics to control *H. pylori* gastric infections [220].

It is known that *H. pylori* uses urease to hydrolyse protein-derived urea available in the human gut lumen in order to synthesise ammonia; up to 10% of the total protein content of the *H. pylori* organism comprises the urease enzyme [221]. The presence of urease is also essential to enable *H. pylori* to colonise the gastric mucosa [222]. This results in partial neutralisation of the low gastric pH. The resultant elevated pH provides the preferred conditions that enable *H. pylori* to thrive (Figure 11). Several pH-sensitive urease inhibitors of varying potency have been identified, and these include ammonia (the product of urease on its substrate), thiols, sulfite, fluoride, green tea-derived epigallocatechin gallate (EGCG), and heavy metals [222].

H. pylori is not the only colonising microbe with urease activity. According to Auron and Brophy in 2012, ureolytic

microbes in the digestive or urinary tracts potentially contribute to hepatic encephalopathy and coma, resulting in hyperammonaemia and brain intoxication [223]. Several other potentially pathogenic urease-positive microbes have been identified; *Klebsiella aerogenes*, *Brevibacterium ammoniagenes*, *Morganella morganii*, *Proteus mirabilis*, *Staphylococcus saprophyticus*, *Escherichia coli*, *Yersinia enterocolitica*, and *Haemophilus influenzae* are among the better-known [222]. Whether SFN is capable of reducing their virulence in humans by urease inhibition is not yet known.

9. Phytochemicals on the Drug Discovery Path

9.1. How do Clinical Trial Data Inform Dose? For a phytochemical to be considered as a therapeutic agent, it must be evaluated using many of the same tools used in pharmaceutical product development. Whereas a pharmaceutical is typically a single molecule, plants are complex multicomponent mixtures; the phytochemical composition of which is not constant due to factors which include inherent agricultural and environmental variability [224].

Of the published SFN research to date, the intervention materials are nonstandard, with some studies using the pure chemical SFN as the intervention material where others use broccoli vegetable, fresh or dried broccoli sprouts; therefore, comparison of clinical trial outcomes becomes more difficult.

Nevertheless, when working with isolated bioactive phytochemicals and whole foods as a source of the same bioactive, the biopharmaceutical processes typically used in pharmaceutical development should equally apply. The LADME principles (liberation, absorption, distribution, metabolism, and excretion) described in connection with

the pharmacokinetics of pharmaceuticals should be equally relevant to phytochemicals [225]. However, such data is seldom available for the more popular phytochemicals used preventively or medicinally [224]. A comprehensive review on this subject by Pferschy-Wenzig and Bauer [224] highlights the many issues that can be underappreciated by consumers who self-medicate on the basis of limited safety and efficacy data.

The literature for SFN indicates that many researchers have addressed the various LADME principles, thereby producing a more extensive database that is useful for interpreting the dose-response.

9.2. Published Clinical Trials. There are currently over 1900 published papers which appear in a PubMed search using the term, “sulforaphane” (PubMed accessed February 4th, 2019). However, there is a limited number of clinical trials utilising either fresh or processed broccoli sprouts (Table 3). Not all trials quantitatively specify the bioactive content of the intervention material. As a result, it is difficult to interpret their findings in a clinical context.

Table 3 illustrates the range of SFN doses used in selected clinical trials where the endpoint is a common human disease or a disease biomarker. Although these trials are of short duration and with small numbers of participants, these data enhance our understanding of the potential of SFN as a clinical intervention. Although dose forms and study populations and endpoints are different across the selected trials, a pattern emerges to show that clinical outcomes are achievable in conditions such as asthma [226] with daily SFN doses of around 18 mg daily and from 27 to 40 mg in type 2 diabetes [24, 227].

A lower SFN dose of around 9–14 mg daily yielded a positive outcome in the autism study by Singh et al. [228], whereas *H. pylori* control was effective with a higher dose of 30 mg SFN daily. Of the available trials, the prostate-specific antigen (PSA) doubling time after radical prostatectomy selected the higher 60 mg daily dose [229].

In considering SFN as a therapeutic intervention, some important questions to be asked are as follows: “What quantity of starting material is needed to achieve a micromolar concentration which generates a significant clinical outcome?” “How can a broccoli sprout raw material be produced which will be consistent in its composition?” and “Is it possible to produce a broccoli sprout raw material that is a practical solution to consumer needs for a SFN-yielding supplement?”

9.3. The Clinician’s Dilemma in Applying Clinical Trial Data. Unlike products categorised by U.S. law as “dietary supplements,” the subgroups of products claiming to be “nutraceutical supplements” are typically standardised for their bioactivity; this may require that one or more bioactives are specified with each batch produced. Of the various available supplements which list a dried broccoli sprout or seed ingredient, the label disclosure is both inconsistent and misleading. Products labelled as “extracts” are manufactured such that GRN is retained as the extract and the myrosinase enzyme needed to synthesise SFN from its glucoraphanin precursor is inactivated [54].

A consumer or a clinician intending to select an available SFN-yielding supplement on the basis of its dose compared with those used in the peer-reviewed published clinical trials has, until very recently, had great difficulty in doing so, given that sprout and seed “extracts” are typically labelled as containing “sulforaphane glucosinolate,” a descriptive commercial name that refers to “glucoraphanin” [236]. Some conversion of GRN to SFN can occur in response to metabolism by the gut microflora; however, the response is inefficient, having been shown to vary “from about 1% to more than 40% of the dose” [237].

Standardisation of label disclosure to remove inconsistency and ambiguity would greatly assist both clinicians and consumers in determining the appropriate daily dose needed to match the doses used in the clinical trials [236].

9.4. Addressing a Conundrum. Because SFN is derived from a commonly consumed vegetable, it is generally considered to lack adverse effects; the safety of broccoli sprouts has been confirmed [238]. Furthermore, a 2018 publication concluded as follows: “it is clear that SFN is a safe and relatively nontoxic chemopreventive agent and exerts anticancer activities through multiple mechanisms, including regulation of Phase I and Phase II drug-metabolising enzymes, anti-inflammatory activity, cell cycle arrest, induction of apoptosis, and the epigenetic regulation on *Nrf2-Keap1*, *cyclins*, and *CDK*” [239].

However, the use of a phytochemical in chemoprevention engages very different biochemical processes when using the same molecule in chemotherapy; the biochemical behaviour of cancer cells and normal cells is very different [240]. As such, it cannot be assumed that SFN as a chemopreventive can be appropriately utilised in the context of chemotherapy where active cancer has been diagnosed.

No discussion of SFN and Nrf2 would be complete without reference to the fact that both Nrf2 activators and Nrf2 inhibitors can be utilised in cancer therapy [241–243]. Cancer cells are able to hijack the Keap1-Nrf2 system via multiple mechanisms leading to enhanced chemo- and radioresistance and proliferation via metabolic reprogramming as well as inhibition of apoptosis [241]. One such mechanism is associated with stimulating the coordinated induction of hepatic Multidrug Resistance Proteins (MRPs) which are adenosine triphosphate-dependent transporters that efflux chemicals out of cells. This ATP-binding cassette family of Phase III detoxification transporters (ABC transporters) [244] is involved in the efflux of numerous endogenous and exogenous chemicals, including chemotherapeutic drugs. MRPs play a key role in cellular protection by removing xenobiotics, metabolites, and endogenous substrates that can accumulate in tissues and lead to toxicity. The activation of the Nrf2 regulatory pathway stimulates the coordinated induction of hepatic MRPs, such that the effective dose of the drug is reduced [245]; this may include chemotherapeutic drugs.

A clinician may then ask whether it is prudent to consider therapies that activate Nrf2 in the context of a cancer diagnosis. James Watson, well-known 1962 Nobel Laureate [246], more recently [247] expressed his concerns about

the potential risks associated with the use of antioxidant therapy in promoting cancer progression. Watson questions whether free radical-destroying antioxidant nutritional supplements may have caused more cancers than they have prevented [247].

In the same year that Watson published his viewpoint, Sporn and Liby suggested that, aside from the extensive literature on the suppression of carcinogenesis by Nrf2 activation, conversely this transcription factor may be oncogenic and cause resistance to chemotherapy [115]. Their opinion article, they say, is aimed at rationalising these conflicting perspectives by critiquing the context dependence of Nrf2 functions and the experimental methods behind these conflicting data. An important new concern they suggest is the finding that common oncogenes, such as *KRAS*, *BRAF*, and *MYC*, all increase the transcription and activity of NRF2, resulting in an increase in cytoprotective activity within the cancer cell [115]. As well, they query the possible effects of Nrf2 polymorphisms, suggesting that enhancement of *NRF2* activity (caused by mutations) can protect tumours from the cytotoxic effects of reactive oxygen species that are induced by chemotherapy or that may be produced endogenously by oncogenic signalling in advanced tumours.

They conclude and rationalise by suggesting that the effect of Nrf2 activation is largely related to the biological time context, stating that Nrf2 activity is desirable (for the host organism) in early stages of tumourigenesis, when the host is seeking to control premalignant carcinogenesis, but is undesirable in later stages of tumourigenesis, when it could make fully malignant cancer cells become resistant to treatment.

A very recent paper [112] highlights this dual role and its implications for Nrf2 activation. It suggests that because Nrf2 can modulate the detoxification pathways, its effect on anti-cancer drugs may lead to chemoresistance and that the switch between a beneficial and a detrimental role for Nrf2 in cancer cells depends on a number of factors which include the tight control of its activity. This poses an obvious dilemma which is already under active discussion and investigation [113, 115]; SFN and other phytochemicals capable of modulating Nrf2 form part of such investigation [112].

Until this dilemma is resolved, clinicians recommending nutraceutical supplements would be wise to avoid coadministration of any nutraceutical supplement whilst the patient is undergoing chemotherapy. Even though “Principles of Care Guidelines” are promoted by an organisation representing such clinicians, it seems clear from the foregoing discussion that there remains insufficient evidence for coadministration of supplements during oncotherapy [248].

That aside, a different line of investigative research has considered whether a role exists for phytochemicals to be utilised in conjunction with chemotherapy. A number of in vitro studies using different cancer cell lines have investigated the potential for SFN (via several mechanisms) to be utilised in conjunction with chemotherapeutic drugs. The goal of such studies is to enable higher doses of the drug to be used before reaching the toxicity threshold of the normal cells [249–255].

In one study as an example of this process, SFN was shown to reduce the toxicity of the chemotherapeutic drug cadmium selenide (CdSe) in human hepatocytes by induction of GSH synthesis at concentrations of 2.5, 5.0, and 10.0 μM SFN, thereby protecting the liver against cytotoxicity and enabling a higher chemotherapeutic dose to be used [250]. Here, SFN’s effect on GSH concentration exhibited a linear dose-response; however, it is unlikely that the higher concentration could be achieved clinically using either diet-derived or supplemental SFN.

Clearly, there is much to be learned before phytochemicals including SFN [256] can be recommended for patients with diagnosed cancers, whether or not the patient is undergoing chemotherapy. Even so, the case for the disease-preventive (including chemopreventive) effects of cruciferous vegetable consumption in general is strong [257, 258] and the last twenty-seven years have witnessed a growing body of evidence to support the roles of SFN in disease prevention, especially given its superior potency as a highly bioavailable Nrf2 activator [259–264].

Albeit limited, the available SFN clinical trial data indicate positive outcomes for a number of common human conditions for which the SFN doses are known. Perhaps future research will more closely focus on its potential effects in patients with diagnosed cancer with a view to resolving the current conundrum.

10. Conclusion

Many decades of research have established strong links between cellular redox and immune imbalances, and the development of chronic disease and biomarkers associated with oxidative stress and inflammation have verified the relationship. However, several large-scale clinical trials to prevent diseases such as T2DM, CVD, and cancer with antioxidant vitamin supplements failed to demonstrate the expected prevention and, in some cases, led to worsening of the biomarkers.

It was not until the discovery of the transcription factor, Nrf2, in 1994 that it became clear that although an enhanced redox balance within the cells was required, the antioxidant vitamins were unable to deliver this. As the understanding of nutrigenomic principles evolved, it became clear that plants contained bioactive phytochemicals that were capable of activating Nrf2; this resulted in the induction of gene expression that targeted a large battery of the genes associated with the core *upstream* cellular defence processes. A key advantage of using phytochemicals to target Nrf2 *upstream* is that a potent Nrf2 activator is capable of inducing hundreds of genes simultaneously.

Of the phytochemicals with Nrf2 inducer capacity, Brassica-derived SFN is the most potent naturally occurring biomolecule known at this time. It is not only a potent Nrf2 inducer but also highly bioavailable so that modest practical doses can produce significant clinical responses. The daily SFN dose found to achieve beneficial outcomes in most of the available clinical trials is around 20–40 mg. With a potent, myrosinase-active whole broccoli sprout supplement, these doses can be attained with just a few capsules daily.

Other Nrf2 activators such as shown in Figure 6 not only lack potency but also lack the bioavailability to be considered as significant intracellular Nrf2 activators. Our understanding of the roles for poorly bioavailable polyphenols in human health is evolving to one more associated with its interactions with the microbiota and the uncertain functions of the metabolites generated by the microbes [55].

Although most of the research on SFN is associated with its ability to activate Nrf2, it exhibits a range of other effects. This review has discussed the way in which another transcription factor, NF- κ B, which is associated with inflammatory pathways is downregulated by SFN. This dual action of SFN is especially intriguing in that Nrf2 and NF- κ B interact via their own “cross talk”.

Infection control is another key activity of the immune system and is closely associated with NF- κ B. In this vein, SFN has been shown to inhibit the *H. pylori* bacterium, a significant gastric cancer risk factor that is prevalent globally. Since pharmaceutical solutions to *H. pylori* eradication are only partially and temporarily effective, the need for a safe, effective therapy is pressing. SFN has been found to inhibit and may possibly even eradicate *H. pylori* in humans via two separate mechanisms.

This review has explored the issues associated with the development of a nutraceutical supplement with significant ability to beneficially influence many of the *upstream* processes associated with core cellular defences; SFN emerges as a potential candidate of this class. The available dose-response evidence is promising, albeit limited so that larger clinical trials will clearly be needed. Even so, the existing data reveal a dose-response that appear to be reasonably consistent by disease state and tissue type and that doses of around 20–40 mg SFN daily can be provided in practical dose form quantities. SFN's primary advantage over many other phytochemicals lies in its comparatively high bioavailability together with its capacity to potently induce Nrf2 target genes.

Has SFN *come of age* as a clinically relevant nutraceutical in the prevention and treatment of chronic disease? Perhaps not just yet; however, the continuing interest in this somewhat novel phytochemical shows no sign of slowing.

Conflicts of Interest

The author is the founder of a company, Cell-Logic Pty Ltd., which manufactures a sulforaphane-yielding raw material and supplement.

References

- [1] S. Yoo, K. Kim, H. Nam, and D. Lee, “Discovering health benefits of phytochemicals with integrated analysis of the molecular network, Chemical Properties and Ethnopharmacological Evidence,” *Nutrients*, vol. 10, no. 8, article 1042, 2018.
- [2] L. Budisan, D. Gulei, O. M. Zanoaga et al., “Dietary intervention by phytochemicals and their role in modulating coding and non-coding genes in cancer,” *International Journal of Molecular Sciences*, vol. 18, no. 6, article 1178, 2017.

- [3] J. Krzyzanowska, A. Czubačka, and W. Oleszek, “Dietary phytochemicals and human health,” in *Bio-Farms for Nutraceuticals*, M. T. Giardi, G. Rea, and B. Berra, Eds., vol. 698 of *Advances in Experimental Medicine and Biology*, pp. 74–98, Springer, Boston, MA, USA, 2010.
- [4] S. Malireddy, S. R. Kotha, J. D. Secor et al., “Phytochemical antioxidants modulate mammalian cellular epigenome: implications in health and disease,” *Antioxidants & Redox Signaling*, vol. 17, no. 2, pp. 327–339, 2012.
- [5] J. A. Barondess, “Scanning the chronic disease terrain: prospects and opportunities,” *Transactions of the American Clinical and Climatological Association*, vol. 125, pp. 45–56, 2014.
- [6] A. Ceriello, “New insights on oxidative stress and diabetic complications may lead to a “causal” antioxidant therapy,” *Diabetes Care*, vol. 26, no. 5, pp. 1589–1596, 2003.
- [7] A. Ceriello and E. Motz, “Is oxidative stress the pathogenic mechanism underlying insulin resistance, diabetes, and cardiovascular disease? The common soil hypothesis revisited,” *Arteriosclerosis, Thrombosis, and Vascular Biology*, vol. 24, no. 5, pp. 816–823, 2004.
- [8] S. Varadharaj, O. J. Kelly, R. N. Khayat, P. S. Kumar, N. Ahmed, and J. L. Zweier, “Role of dietary antioxidants in the preservation of vascular function and the modulation of health and disease,” *Frontiers in Cardiovascular Medicine*, vol. 4, p. 64, 2017.
- [9] M. Brownlee, “The pathobiology of diabetic complications: a unifying mechanism,” *Diabetes*, vol. 54, no. 6, pp. 1615–1625, 2005.
- [10] A. Ceriello, “Controlling oxidative stress as a novel molecular approach to protecting the vascular wall in diabetes,” *Current Opinion in Lipidology*, vol. 17, no. 5, pp. 510–518, 2006.
- [11] A. Ceriello, “Oxidative stress and diabetes-associated complications,” *Endocrine Practice*, vol. 12, Supplement 1, pp. 60–62, 2006.
- [12] I. A. Kirpich, D. Parajuli, and C. J. McClain, “The gut microbiome in NAFLD and ALD,” *Clinical Liver Disease*, vol. 6, no. 3, pp. 55–58, 2015.
- [13] L. A. Velloso, F. Folli, and M. J. Saad, “TLR4 at the crossroads of nutrients, gut microbiota, and metabolic inflammation,” *Endocrine Reviews*, vol. 36, no. 3, pp. 245–271, 2015.
- [14] E. Eren, K. U. Tufekci, K. B. Isci, B. Tastan, K. Genc, and S. Genc, “Sulforaphane inhibits lipopolysaccharide-induced inflammation, cytotoxicity, oxidative stress, and miR-155 expression and switches to Mox phenotype through activating extracellular signal-regulated kinase 1/2–nuclear factor erythroid 2-related factor 2/antioxidant response element pathway in murine microglial cells,” *Frontiers in Immunology*, vol. 9, p. 36, 2018.
- [15] L. Subedi, J. H. Lee, S. Yumnam, E. Ji, and S. Y. Kim, “Anti-inflammatory effect of sulforaphane on LPS-activated microglia potentially through JNK/AP-1/NF- κ B inhibition and Nrf2/HO-1 activation,” *Cells*, vol. 8, no. 2, p. 194, 2019.
- [16] G. Bjelakovic, D. Nikolova, L. L. Gluud, R. G. Simonetti, and C. Gluud, “Mortality in randomized trials of antioxidant supplements for primary and secondary prevention: systematic review and meta-analysis,” *JAMA*, vol. 297, no. 8, pp. 842–857, 2007.
- [17] L. Franzini, D. Ardigo, and I. Zavaroni, “Dietary antioxidants and glucose metabolism,” *Current Opinion in Clinical Nutrition and Metabolic Care*, vol. 11, no. 4, pp. 471–476, 2008.
- [18] Y. Song, N. R. Cook, C. M. Albert, M. Van Denburgh, and J. E. Manson, “Effects of vitamins C and E and β -carotene

- on the risk of type 2 diabetes in women at high risk of cardiovascular disease: a randomized controlled trial," *The American Journal of Clinical Nutrition*, vol. 90, no. 2, pp. 429–437, 2009.
- [19] H. Priemé, S. Loft, K. Nyssönen, J. T. Salonen, and H. E. Poulsen, "No effect of supplementation with vitamin E, ascorbic acid, or coenzyme Q10 on oxidative DNA damage estimated by 8-oxo-7,8-dihydro-2'-deoxyguanosine excretion in smokers," *The American Journal of Clinical Nutrition*, vol. 65, no. 2, pp. 503–507, 1997.
- [20] Y. C. Chang and L. M. Chuang, "The role of oxidative stress in the pathogenesis of type 2 diabetes: from molecular mechanism to clinical implication," *American Journal of Translational Research*, vol. 2, no. 3, pp. 316–331, 2010.
- [21] C. Hart, R. Cohen, M. Norwood, and J. Stebbing, "The emerging harm of antioxidants in carcinogenesis," *Future Oncology*, vol. 8, no. 5, pp. 535–548, 2012.
- [22] G. Bjelakovic, D. Nikolova, L. L. Gluud, R. G. Simonetti, and C. Gluud, "Antioxidant supplements for prevention of mortality in healthy participants and patients with various diseases," *Cochrane Database of Systematic Reviews*, no. 3, article CD007176, 2012.
- [23] G. Bjelakovic and C. Gluud, "Vitamin and mineral supplement use in relation to all-cause mortality in the Iowa Women's Health Study: comment on "Dietary Supplements and Mortality Rate in Older Women"," *Archives of Internal Medicine*, vol. 171, no. 18, pp. 1633–1634, 2011.
- [24] Z. Bahadoran, P. Mirmiran, F. Hosseinpour-Niazi, M. Hedayati, S. Hosseinpour-Niazi, and F. Azizi, "Broccoli sprouts reduce oxidative stress in type 2 diabetes: a randomized double-blind clinical trial," *European Journal of Clinical Nutrition*, vol. 65, no. 8, pp. 972–977, 2011.
- [25] M. Murashima, S. Watanabe, X. G. Zhuo, M. Uehara, and A. Kurashige, "Phase 1 study of multiple biomarkers for metabolism and oxidative stress after one-week intake of broccoli sprouts," *BioFactors*, vol. 22, no. 1–4, pp. 271–275, 2004.
- [26] T. W. Kensler, J. G. Chen, P. A. Egner et al., "Effects of glucosinolate-rich broccoli sprouts on urinary levels of aflatoxin-DNA adducts and phenanthrene tetraols in a randomized clinical trial in He Zuo township, Qidong, People's Republic of China," *Cancer Epidemiology, Biomarkers & Prevention*, vol. 14, no. 11, pp. 2605–2613, 2005.
- [27] M. C. Myzak, P. A. Karplus, F.-L. Chung, and R. H. Dashwood, "A novel mechanism of chemoprotection by sulforaphane: inhibition of histone deacetylase," *Cancer Research*, vol. 64, no. 16, pp. 5767–5774, 2004.
- [28] C. Fimognari and P. Hrelia, "Sulforaphane as a promising molecule for fighting cancer," *Mutation Research/Reviews in Mutation Research*, vol. 635, no. 2–3, pp. 90–104, 2007.
- [29] M. A. Riedl, A. Saxon, and D. Diaz-Sanchez, "Oral sulforaphane increases Phase II antioxidant enzymes in the human upper airway," *Clinical Immunology*, vol. 130, no. 3, pp. 244–251, 2009.
- [30] C. Livingstone and J. Davis, "Review: targeting therapeutics against glutathione depletion in diabetes and its complications," *The British Journal of Diabetes & Vascular Disease*, vol. 7, no. 6, pp. 258–265, 2007.
- [31] D. Harman, "Free radical theory of aging," *Mutation Research/DNAging*, vol. 275, no. 3–6, pp. 257–266, 1992.
- [32] S. Chambial, S. Dwivedi, K. K. Shukla, P. J. John, and P. Sharma, "Vitamin C in disease prevention and cure: an overview," *Indian Journal of Clinical Biochemistry*, vol. 28, no. 4, pp. 314–328, 2013.
- [33] M. J. Howes and M. S. Simmonds, "The role of phytochemicals as micronutrients in health and disease," *Current Opinion in Clinical Nutrition and Metabolic Care*, vol. 17, no. 6, pp. 558–566, 2014.
- [34] D. E. Stevenson and R. D. Hurst, "Polyphenolic phytochemicals – just antioxidants or much more?," *Cellular and Molecular Life Sciences*, vol. 64, no. 22, pp. 2900–2916, 2007.
- [35] V. Calabrese, C. Cornelius, A. T. Dinkova-Kostova et al., "Cellular stress responses, hormetic phytochemicals and vitamins in aging and longevity," *Biochimica et Biophysica Acta (BBA) - Molecular Basis of Disease*, vol. 1822, no. 5, pp. 753–783, 2012.
- [36] R. H. Liu, "Potential synergy of phytochemicals in cancer prevention: mechanism of action," *The Journal of Nutrition*, vol. 134, no. 12, pp. 3479S–3485S, 2004.
- [37] P. Ghezzi, V. Jaquet, F. Marcucci, and H. Schmidt, "The oxidative stress theory of disease: levels of evidence and epistemological aspects," *British Journal of Pharmacology*, vol. 174, no. 12, pp. 1784–1796, 2017.
- [38] A. Dandekar, R. Mendez, and K. Zhang, "Cross talk between ER stress, oxidative stress, and inflammation in health and disease," *Methods in Molecular Biology*, vol. 1292, pp. 205–214, 2015.
- [39] G. Bjelakovic and C. Gluud, "Surviving antioxidant supplements," *JNCI: Journal of the National Cancer Institute*, vol. 99, no. 10, pp. 742–743, 2007.
- [40] G. Bjelakovic, L. L. Gluud, D. Nikolova, M. Bjelakovic, A. Nagorni, and C. Gluud, "Antioxidant supplements for liver diseases," *Cochrane Database of Systematic Reviews*, no. 3, article CD007749, 2011.
- [41] G. Bjelakovic, A. Nagorni, D. Nikolova, R. G. Simonetti, M. Bjelakovic, and C. Gluud, "Meta-analysis: antioxidant supplements for primary and secondary prevention of colorectal adenoma," *Alimentary Pharmacology and Therapeutics*, vol. 24, no. 2, pp. 281–291, 2006.
- [42] G. Bjelakovic, D. Nikolova, L. L. Gluud, R. G. Simonetti, and C. Gluud, "Antioxidant supplements for prevention of mortality in healthy participants and patients with various diseases," *Cochrane Database of Systematic Reviews*, no. 2, article CD007176, 2012.
- [43] G. Bjelakovic, D. Nikolova, R. G. Simonetti, and C. Gluud, "Antioxidant supplements for preventing gastrointestinal cancers," *Cochrane Database of Systematic Reviews*, vol. 4, article CD004183, 2004.
- [44] G. Bjelakovic, D. Nikolova, R. G. Simonetti, and C. Gluud, "Antioxidant supplements for prevention of gastrointestinal cancers: a systematic review and meta-analysis," *The Lancet*, vol. 364, no. 9441, pp. 1219–1228, 2004.
- [45] G. Bjelakovic, D. Nikolova, R. G. Simonetti, and C. Gluud, "Systematic review: primary and secondary prevention of gastrointestinal cancers with antioxidant supplements," *Alimentary Pharmacology & Therapeutics*, vol. 28, no. 6, pp. 689–703, 2008.
- [46] G. Bjelakovic, D. Nikolova, R. G. Simonetti, and C. Gluud, "Antioxidant supplements for preventing gastrointestinal cancers," *Cochrane Database of Systematic Reviews*, no. 3, article CD004183, 2008.
- [47] J. Kaput, J. Noble, B. Hatipoglu, K. Kohrs, K. Dawson, and A. Bartholomew, "Application of nutrigenomic concepts to

- Type 2 diabetes mellitus," *Nutrition, Metabolism, and Cardiovascular Diseases*, vol. 17, no. 2, pp. 89–103, 2007.
- [48] J. Higdon, *An Evidence-Based Approach to Dietary Phytochemicals*, Thieme Medical Publishers, New York, 2007.
- [49] M. Muller and S. Kersten, "Nutrigenomics: goals and strategies," *Nature Reviews Genetics*, vol. 4, no. 4, pp. 315–322, 2003.
- [50] S. N. B. Selby-Pham, R. B. Miller, K. Howell, F. Dunshea, and L. E. Bennett, "Physicochemical properties of dietary phytochemicals can predict their passive absorption in the human small intestine," *Scientific Reports*, vol. 7, no. 1, pp. 1931–1931, 2017.
- [51] D. Vauzour, A. Rodriguez-Mateos, G. Corona, M. J. Oruna-Concha, and J. P. Spencer, "Polyphenols and human health: prevention of disease and mechanisms of action," *Nutrients*, vol. 2, no. 11, pp. 1106–1131, 2010.
- [52] P. E. Milbury, J. A. Vita, and J. B. Blumberg, "Anthocyanins are bioavailable in humans following an acute dose of cranberry juice," *The Journal of Nutrition*, vol. 140, no. 6, pp. 1099–1104, 2010.
- [53] M. D'Archivio, C. Filesi, R. Vari, B. Scaccocchio, and R. Masella, "Bioavailability of the polyphenols: status and controversies," *International Journal of Molecular Sciences*, vol. 11, no. 4, pp. 1321–1342, 2010.
- [54] C. A. Houghton, R. G. Fassett, and J. S. Coombes, "Sulforaphane and other nutrigenomic Nrf2 activators: can the clinician's expectation be matched by the reality?," *Oxidative Medicine and Cellular Longevity*, vol. 2016, Article ID 7857186, 17 pages, 2016.
- [55] T. Ozdal, D. A. Sela, J. Xiao, D. Boyacioglu, F. Chen, and E. Capanoglu, "The reciprocal interactions between polyphenols and gut microbiota and effects on bioaccessibility," *Nutrients*, vol. 8, no. 2, pp. 78–78, 2016.
- [56] F. Poti, D. Santi, G. Spaggiari, F. Zimetti, and I. Zanotti, "Polyphenol health effects on cardiovascular and neurodegenerative disorders: a review and meta-analysis," *International Journal of Molecular Sciences*, vol. 20, no. 2, 2019.
- [57] N. Hanlon, N. Coldham, A. Gielbert et al., "Absolute bioavailability and dose-dependent pharmacokinetic behaviour of dietary doses of the chemopreventive isothiocyanate sulforaphane in rat," *The British Journal of Nutrition*, vol. 99, no. 3, pp. 559–564, 2008.
- [58] R. K. Thimmulappa, K. H. Mai, S. Srisuma, T. W. Kensler, M. Yamamoto, and S. Biswal, "Identification of Nrf2-regulated genes induced by the chemopreventive agent sulforaphane by oligonucleotide microarray," *Cancer Research*, vol. 62, no. 18, pp. 5196–5203, 2002.
- [59] N. Hanlon, C. L. Poynton, N. Coldham, M. J. Sauer, and C. Ioannides, "The aliphatic isothiocyanates erucin and sulforaphane do not effectively up-regulate NAD(P)H:quinone oxidoreductase (NQO1) in human liver compared with rat," *Molecular Nutrition & Food Research*, vol. 53, no. 7, pp. 836–844, 2009.
- [60] S. J. Franklin, S. E. Dickinson, K. L. Karlage, G. T. Bowden, and P. B. Myrdal, "Stability of sulforaphane for topical formulation," *Drug Development and Industrial Pharmacy*, vol. 40, no. 4, pp. 494–502, 2014.
- [61] D. N. Brooke, A. J. Dobbs, and N. Williams, "Octanol: Water partition coefficients (P): Measurement, estimation, and interpretation, particularly for chemicals with $P > 10^5$," *Ecotoxicology and Environmental Safety*, vol. 11, no. 3, pp. 251–260, 1986.
- [62] N. Juge, R. F. Mithen, and M. Traka, "Molecular basis for chemoprevention by sulforaphane: a comprehensive review," *Cellular and Molecular Life Sciences*, vol. 64, pp. 1105–1127, 2007.
- [63] J. D. Wardyn, A. H. Ponsford, and C. M. Sanderson, "Dissecting molecular cross-talk between Nrf2 and NF- κ B response pathways," *Biochemical Society Transactions*, vol. 43, no. 4, pp. 621–626, 2015.
- [64] J. W. Fahey and P. Talalay, "Antioxidant functions of sulforaphane: a potent inducer of Phase II detoxication enzymes," *Food and Chemical Toxicology*, vol. 37, no. 9-10, pp. 973–979, 1999.
- [65] A. T. Dinkova-Kostova and P. Talalay, "Direct and indirect antioxidant properties of inducers of cytoprotective proteins," *Molecular Nutrition & Food Research*, vol. 52, no. S1, pp. S128–S138, 2008.
- [66] P. Talalay, "Chemoprotection against cancer by induction of phase 2 enzymes," *BioFactors*, vol. 12, no. 1-4, pp. 5–11, 2000.
- [67] T. W. Kensler, N. Wakabayashi, and S. Biswal, "Cell survival responses to environmental stresses via the Keap1-Nrf2-ARE pathway," *Annual Review of Pharmacology and Toxicology*, vol. 47, no. 1, pp. 89–116, 2007.
- [68] W. D. Holtzclaw, A. T. Dinkova-Kostova, and P. Talalay, "Protection against electrophile and oxidative stress by induction of phase 2 genes: the quest for the elusive sensor that responds to inducers," *Advances in Enzyme Regulation*, vol. 44, no. 1, pp. 335–367, 2004.
- [69] X. Cheng, R. C. Siow, and G. E. Mann, "Impaired redox signaling and antioxidant gene expression in endothelial cells in diabetes: a role for mitochondria and the nuclear factor-E2-related factor 2-Kelch-like ECH-associated protein 1 defense pathway," *Antioxidants & Redox Signaling*, vol. 14, no. 3, pp. 469–487, 2011.
- [70] C. Manach, A. Scalbert, C. Morand, C. Remesy, and L. Jimenez, "Polyphenols: food sources and bioavailability," *The American Journal of Clinical Nutrition*, vol. 79, no. 5, pp. 727–747, 2004.
- [71] S. Joko, M. Watanabe, H. Fuda et al., "Comparison of chemical structures and cytoprotection abilities between direct and indirect antioxidants," *Journal of Functional Foods*, vol. 35, pp. 245–255, 2017.
- [72] H. Teng and L. Chen, "Polyphenols and bioavailability: an update," *Critical Reviews in Food Science and Nutrition*, vol. 59, no. 13, pp. 2040–2051, 2019.
- [73] P. Moi, K. Chan, I. Asunis, A. Cao, and Y. W. Kan, "Isolation of NF-E2-related factor 2 (Nrf2), a NF-E2-like basic leucine zipper transcriptional activator that binds to the tandem NF-E2/AP1 repeat of the beta-globin locus control region," *Proceedings of the National Academy of Sciences of the United States of America*, vol. 91, no. 21, pp. 9926–9930, 1994.
- [74] M. K. Kwak, N. Wakabayashi, and T. W. Kensler, "Chemoprevention through the Keap1-Nrf2 signaling pathway by phase 2 enzyme inducers," *Mutation Research/Fundamental and Molecular Mechanisms of Mutagenesis*, vol. 555, no. 1-2, pp. 133–148, 2004.
- [75] Y. Shinkai, D. Sumi, I. Fukami, T. Ishii, and Y. Kumagai, "Sulforaphane, an activator of Nrf2, suppresses cellular accumulation of arsenic and its cytotoxicity in primary mouse hepatocytes," *FEBS Letters*, vol. 580, no. 7, pp. 1771–1774, 2006.

- [76] S. Vomund, A. Schäfer, M. J. Parnham, B. Brüne, and A. von Knethen, "Nrf2, the master regulator of anti-oxidative responses," *International Journal of Molecular Sciences*, vol. 18, no. 12, p. 2772, 2017.
- [77] A. L. Egger and S. N. Savinov, "Chemical and biological mechanisms of phytochemical activation of Nrf2 and importance in disease prevention," *Recent Advances in Phytochemistry*, vol. 43, pp. 121–155, 2013.
- [78] H.-C. Hung, K. J. Joshipura, R. Jiang et al., "Fruit and vegetable intake and risk of major chronic disease," *Journal of the National Cancer Institute*, vol. 96, no. 21, pp. 1577–1584, 2004.
- [79] Y. Zhang, P. Talalay, C. G. Cho, and G. H. Posner, "A major inducer of anticarcinogenic protective enzymes from broccoli: isolation and elucidation of structure," *Proceedings of the National Academy of Sciences of the United States of America*, vol. 89, no. 6, pp. 2399–2403, 1992.
- [80] H. J. Prochaska, A. B. Santamaria, and P. Talalay, "Rapid detection of inducers of enzymes that protect against carcinogens," *Proceedings of the National Academy of Sciences of the United States of America*, vol. 89, no. 6, pp. 2394–2398, 1992.
- [81] J. W. Fahey, Y. Zhang, and P. Talalay, "Broccoli sprouts: an exceptionally rich source of inducers of enzymes that protect against chemical carcinogens," *Proceedings of the National Academy of Sciences of the United States of America*, vol. 94, no. 19, pp. 10367–10372, 1997.
- [82] V. V. Lyakhovich, V. A. Vavilin, N. K. Zenkov, and E. B. Menshchikova, "Active defense under oxidative stress. The antioxidant responsive element," *Biochemistry (Moscow)*, vol. 71, no. 9, pp. 962–974, 2006.
- [83] R. Brigelius-Flohé and A. Banning, "Part of the Series: From dietary antioxidants to regulators in cellular signaling and gene regulation," *Free Radical Research*, vol. 40, no. 8, pp. 775–787, 2006.
- [84] A. Seow, C.-Y. Shi, F.-L. Chung et al., "Urinary total isothiocyanate (ITC) in a population-based sample of middle-aged and older Chinese in Singapore: relationship with dietary total ITC and glutathione S-transferase M1/T1/P1 genotypes," *Cancer Epidemiology, Biomarkers & Prevention*, vol. 7, no. 9, pp. 775–781, 1998.
- [85] J. W. Lampe and S. Peterson, "Brassica, biotransformation and cancer risk: genetic polymorphisms alter the preventive effects of cruciferous vegetables," *The Journal of Nutrition*, vol. 132, no. 10, pp. 2991–2994, 2002.
- [86] A. T. Dinkova-Kostova, J. W. Fahey, R. V. Kostov, and T. W. Kensler, "KEAP1 and done? Targeting the NRF2 pathway with sulforaphane," *Trends in Food Science & Technology*, vol. 69, Part B, pp. 257–269, 2017.
- [87] S. A. McNaughton and G. C. Marks, "Development of a food composition database for the estimation of dietary intakes of glucosinolates, the biologically active constituents of cruciferous vegetables," *The British Journal of Nutrition*, vol. 90, no. 3, pp. 687–697, 2003.
- [88] B. S. Cornblatt, L. Ye, A. T. Dinkova-Kostova et al., "Preclinical and clinical evaluation of sulforaphane for chemoprevention in the breast," *Carcinogenesis*, vol. 28, no. 7, pp. 1485–1490, 2007.
- [89] L. Ye, A. T. Dinkova-Kostova, K. L. Wade, Y. Zhang, T. A. Shapiro, and P. Talalay, "Quantitative determination of dithiocarbamates in human plasma, serum, erythrocytes and urine: pharmacokinetics of broccoli sprout isothiocyanates in humans," *Clinica Chimica Acta*, vol. 316, no. 1–2, pp. 43–53, 2002.
- [90] C. Manach, G. Williamson, C. Morand, A. Scalbert, and C. Rémésy, "Bioavailability and bioefficacy of polyphenols in humans. I. Review of 97 bioavailability studies," *The American Journal of Clinical Nutrition*, vol. 81, no. 1, pp. 230S–242S, 2005.
- [91] J. M. Lee and J. A. Johnson, "An important role of Nrf2-ARE pathway in the cellular defense mechanism," *BMB Reports*, vol. 37, no. 2, pp. 139–143, 2004.
- [92] Y. J. Surh, J. K. Kundu, and H. K. Na, "Nrf2 as a master redox switch in turning on the cellular signaling involved in the induction of cytoprotective genes by some chemopreventive phytochemicals," *Planta Medica*, vol. 74, no. 13, pp. 1526–1539, 2008.
- [93] K. N. Lewis, J. Mele, J. D. Hayes, and R. Buffenstein, "Nrf2, a guardian of healthspan and gatekeeper of species longevity," *Integrative and Comparative Biology*, vol. 50, no. 5, pp. 829–843, 2010.
- [94] A. Rahal, A. Kumar, V. Singh et al., "Oxidative stress, prooxidants, and antioxidants: the interplay," *BioMed Research International*, vol. 2014, Article ID 761264, 19 pages, 2014.
- [95] A. T. Dinkova-Kostova, W. D. Holtzclaw, and T. W. Kensler, "The role of Keap1 in cellular protective responses," *Chemical Research in Toxicology*, vol. 18, no. 12, pp. 1779–1791, 2005.
- [96] T. Prester, W. D. Holtzclaw, Y. Zhang, and P. Talalay, "Chemical and molecular regulation of enzymes that detoxify carcinogens," *Proceedings of the National Academy of Sciences of the United States of America*, vol. 90, no. 7, pp. 2965–2969, 1993.
- [97] M. Zhang, C. An, Y. Gao, R. K. Leak, J. Chen, and F. Zhang, "Emerging roles of Nrf2 and phase II antioxidant enzymes in neuroprotection," *Progress in Neurobiology*, vol. 100, pp. 30–47, 2013.
- [98] M. S. Yates, Q. T. Tran, P. M. Dolan et al., "Genetic versus chemoprotective activation of Nrf2 signaling: overlapping yet distinct gene expression profiles between Keap1 knockout and triterpenoid-treated mice," *Carcinogenesis*, vol. 30, no. 6, pp. 1024–1031, 2009.
- [99] A. T. Dinkova-Kostova, W. D. Holtzclaw, R. N. Cole et al., "Direct evidence that sulfhydryl groups of Keap1 are the sensors regulating induction of phase 2 enzymes that protect against carcinogens and oxidants," *Proceedings of the National Academy of Sciences of the United States of America*, vol. 99, no. 18, pp. 11908–11913, 2002.
- [100] R. Munday, Y. Zhang, J. D. Paonessa, C. M. Munday, A. L. Wilkins, and J. Babu, "Synthesis, Biological Evaluation, and Structure–Activity Relationships of Dithiolethiones as Inducers of Cytoprotective Phase 2 Enzymes," *Journal of Medicinal Chemistry*, vol. 53, no. 12, pp. 4761–4767, 2010.
- [101] R. Munday, Y. Zhang, C. M. Munday, and J. Li, "Structure–activity relationships in the induction of Phase II enzymes by derivatives of 3H-1,2-dithiole-3-thione in rats," *Chemico-Biological Interactions*, vol. 160, no. 2, pp. 115–122, 2006.
- [102] Y. Zhang and R. Munday, "Dithiolethiones for cancer chemoprevention: where do we stand?," *Molecular Cancer Therapeutics*, vol. 7, no. 11, pp. 3470–3479, 2008.
- [103] L. M. Aleksunes, M. J. Goedken, C. E. Rockwell, J. Thomale, J. E. Manautou, and C. D. Klaassen, "Transcriptional regulation of renal cytoprotective genes by Nrf2 and its potential

- use as a therapeutic target to mitigate cisplatin-induced nephrotoxicity," *The Journal of Pharmacology and Experimental Therapeutics*, vol. 335, no. 1, pp. 2–12, 2010.
- [104] H. Liu and P. Talalay, "Relevance of anti-inflammatory and antioxidant activities of exemestane and synergism with sulforaphane for disease prevention," *Proceedings of the National Academy of Sciences of the United States of America*, vol. 110, no. 47, pp. 19065–19070, 2013.
- [105] M. C. Myzak and R. H. Dashwood, "Histone deacetylases as targets for dietary cancer preventive agents: lessons learned with butyrate, diallyl disulfide, and sulforaphane," *Current Drug Targets*, vol. 7, no. 4, pp. 443–452, 2006.
- [106] R. H. Dashwood, M. C. Myzak, and E. Ho, "Dietary HDAC inhibitors: time to rethink weak ligands in cancer chemoprevention?," *Carcinogenesis*, vol. 27, no. 2, pp. 344–349, 2006.
- [107] J. Huang, C. Plass, and C. Gerhauser, "Cancer chemoprevention by targeting the epigenome," *Current Drug Targets*, vol. 12, no. 13, pp. 1925–1956, 2011.
- [108] J. H. Park, Y. Yoo, and Y. J. Park, "Epigenetics: linking nutrition to molecular mechanisms in aging," *Preventive Nutrition and Food Science*, vol. 22, no. 2, pp. 81–89, 2017.
- [109] S. L. Martin, R. Kala, and T. O. Tollefsbol, "Mechanisms for the Inhibition of Colon Cancer Cells by Sulforaphane through Epigenetic Modulation of MicroRNA-21 and Human Telomerase Reverse Transcriptase (hTERT) Down-regulation," *Current Cancer Drug Targets*, vol. 18, no. 1, 2017.
- [110] A. Kaufman-Szymczyk, G. Majewski, K. Lubecka-Pietruszewska, and K. Fabianowska-Majewska, "The role of sulforaphane in epigenetic mechanisms, including interdependence between histone modification and DNA methylation," *International Journal of Molecular Sciences*, vol. 16, no. 12, pp. 29732–29743, 2015.
- [111] A. J. Levine and S. L. Berger, "The interplay between epigenetic changes and the p53 protein in stem cells," *Genes & Development*, vol. 31, no. 12, pp. 1195–1201, 2017.
- [112] E. Catanzaro, C. Calcabrini, E. Turrini, P. Sestili, and C. Fimognari, "Nrf2: a potential therapeutic target for naturally occurring anticancer drugs?," *Expert Opinion on Therapeutic Targets*, vol. 21, no. 8, pp. 781–793, 2017.
- [113] X. Bai, Y. Chen, X. Hou, M. Huang, and J. Jin, "Emerging role of NRF2 in chemoresistance by regulating drug-metabolizing enzymes and efflux transporters," *Drug Metabolism Reviews*, vol. 48, no. 4, pp. 541–567, 2016.
- [114] T. W. Kensler and N. Wakabayashi, "Nrf2: friend or foe for chemoprevention?," *Carcinogenesis*, vol. 31, no. 1, pp. 90–99, 2010.
- [115] M. B. Sporn and K. T. Liby, "NRF2 and cancer: the good, the bad and the importance of context," *Nature Reviews Cancer*, vol. 12, no. 8, pp. 564–571, 2012.
- [116] B. N. Chorley, M. R. Campbell, X. Wang et al., "Identification of novel NRF2-regulated genes by ChIP-Seq: influence on retinoid X receptor alpha," *Nucleic Acids Research*, vol. 40, no. 15, pp. 7416–7429, 2012.
- [117] Y. Zhang and L. Tang, "Discovery and development of sulforaphane as a cancer chemopreventive phytochemical," *Acta Pharmacologica Sinica*, vol. 28, no. 9, pp. 1343–1354, 2007.
- [118] S. V. Singh, S. K. Srivastava, S. Choi et al., "Sulforaphane-induced cell death in human prostate cancer cells is initiated by reactive oxygen species," *The Journal of Biological Chemistry*, vol. 280, no. 20, pp. 19911–19924, 2005.
- [119] A. Pastore, G. Federici, E. Bertini, and F. Piemonte, "Analysis of glutathione: implication in redox and detoxification," *Clinica Chimica Acta*, vol. 333, no. 1, pp. 19–39, 2003.
- [120] M. C. Myzak, P. A. Karplus, F. L. Chung, and R. H. Dashwood, "A Novel Mechanism of Chemoprotection by Sulforaphane," *Cancer Research*, vol. 64, no. 16, pp. 5767–5774, 2004.
- [121] H. Steinkellner, S. Rabot, C. Freywald et al., "Effects of cruciferous vegetables and their constituents on drug metabolizing enzymes involved in the bioactivation of DNA-reactive dietary carcinogens," *Mutation Research/Fundamental and Molecular Mechanisms of Mutagenesis*, vol. 480–481, pp. 285–297, 2001.
- [122] M. C. Myzak, K. Hardin, R. Wang, R. H. Dashwood, and E. Ho, "Sulforaphane inhibits histone deacetylase activity in BPH-1, LnCaP and PC-3 prostate epithelial cells," *Carcinogenesis*, vol. 27, no. 4, pp. 811–819, 2006.
- [123] C. Fimognari, F. Berti, G. Cantelli-Forti, and P. Hrelia, "Effect of sulforaphane on micronucleus induction in cultured human lymphocytes by four different mutagens," *Environmental and Molecular Mutagenesis*, vol. 46, no. 4, pp. 260–267, 2005.
- [124] L. Tang, Y. Zhang, H. E. Jobson et al., "Potent activation of mitochondria-mediated apoptosis and arrest in S and M phases of cancer cells by a broccoli sprout extract," *Molecular Cancer Therapeutics*, vol. 5, no. 4, pp. 935–944, 2006.
- [125] S. A. Ritz, J. Wan, and D. Diaz-Sanchez, "Sulforaphane-stimulated phase II enzyme induction inhibits cytokine production by airway epithelial cells stimulated with diesel extract," *American Journal of Physiology-Lung Cellular and Molecular Physiology*, vol. 292, no. 1, pp. L33–L39, 2007.
- [126] S. Mukherjee, H. Gangopadhyay, and D. K. Das, "Broccoli: a unique vegetable that protects mammalian hearts through the redox cycling of the thioredoxin superfamily," *Journal of Agricultural and Food Chemistry*, vol. 56, no. 2, pp. 609–617, 2008.
- [127] J. W. Fahey, X. Haristoy, P. M. Dolan et al., "Sulforaphane inhibits extracellular, intracellular, and antibiotic-resistant strains of *Helicobacter pylori* and prevents benzo[a]pyrene-induced stomach tumors," *Proceedings of the National Academy of Sciences of the United States of America*, vol. 99, no. 11, pp. 7610–7615, 2002.
- [128] J. M. Han, Y. J. Lee, S. Y. Lee et al., "Protective effect of sulforaphane against dopaminergic cell death," *The Journal of Pharmacology and Experimental Therapeutics*, vol. 321, no. 1, pp. 249–256, 2007.
- [129] C. Fimognari, M. Nusse, F. Berti, R. Iori, G. Cantelli-Forti, and P. Hrelia, "Cyclin D3 and p53 mediate sulforaphane-induced cell cycle delay and apoptosis in non-transformed human T lymphocytes," *Cellular and Molecular Life Sciences*, vol. 59, no. 11, pp. 2004–2012, 2002.
- [130] Y.-J. Lee and S.-H. Lee, "Sulforaphane Induces Antioxidative and Antiproliferative Responses by Generating Reactive Oxygen Species in Human Bronchial Epithelial BEAS-2B Cells," *Journal of Korean Medical Science*, vol. 26, no. 11, pp. 1474–1482, 2011.
- [131] T. Hisada, M. Salmon, Y. Nasuhara, and K. F. Chung, "Involvement of haemoxygenase-1 in ozone-induced airway inflammation and hyperresponsiveness," *European Journal of Pharmacology*, vol. 399, no. 2–3, pp. 229–234, 2000.
- [132] S. Murata, T. Matsuda, S. Kiguchi, M. Kobayashi, K. Cho, and K. Okuyama, "Effects of long term administration of KUR-

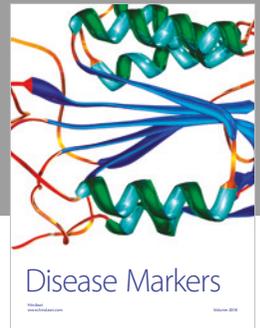
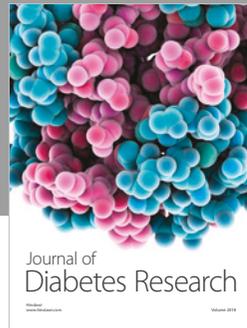
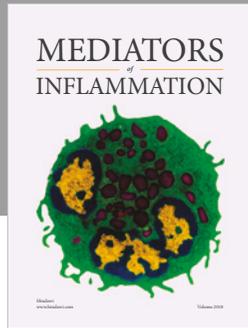
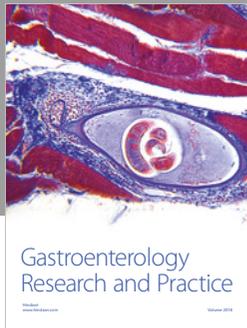
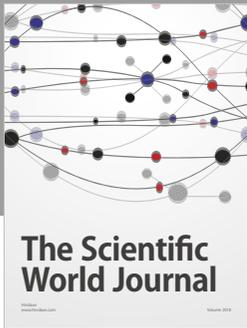
- 1246, a selective beta2-adrenoceptor agonist, on pregnant sheep and their fetuses." *BJOG: An International Journal of Obstetrics and Gynaecology*, vol. 112, no. 1, pp. 69–74, 2005.
- [133] X. L. Chen and C. Kunsch, "Induction of Cytoprotective Genes Through Nrf2 / Antioxidant Response Element Pathway: A New Therapeutic Approach for the Treatment of Inflammatory Diseases," *Current Pharmaceutical Design*, vol. 10, no. 8, pp. 879–891, 2004.
- [134] R. Hu, C. Xu, G. Shen et al., "Gene expression profiles induced by cancer chemopreventive isothiocyanate sulforaphane in the liver of C57BL/6J mice and C57BL/6J/Nrf2 (–/–) mice," *Cancer Letters*, vol. 243, no. 2, pp. 170–192, 2006.
- [135] Y. Zhang and P. Talalay, "Mechanism of differential potencies of isothiocyanates as inducers of anticarcinogenic Phase 2 enzymes," *Cancer Research*, vol. 58, no. 20, pp. 4632–4639, 1998.
- [136] J. D. Hayes and M. McMahon, "NRF2 and KEAP1 mutations: permanent activation of an adaptive response in cancer," *Trends in Biochemical Sciences*, vol. 34, no. 4, pp. 176–188, 2009.
- [137] V. Calabrese, J. Giordano, M. Ruggieri et al., "Hormesis, cellular stress response, and redox homeostasis in autism spectrum disorders," *Journal of Neuroscience Research*, vol. 94, no. 12, pp. 1488–1498, 2016.
- [138] J. D. Brooks, V. G. Paton, and G. Vidanes, "Potent induction of phase 2 enzymes in human prostate cells by sulforaphane," *Cancer Epidemiology, Biomarkers & Prevention*, vol. 10, no. 9, pp. 949–954, 2001.
- [139] D. Vauzour, M. Buonfiglio, G. Corona et al., "Sulforaphane protects cortical neurons against 5-S-cysteinyl-dopamine-induced toxicity through the activation of ERK1/2, Nrf-2 and the upregulation of detoxification enzymes," *Molecular Nutrition & Food Research*, vol. 54, no. 4, pp. 532–542, 2010.
- [140] H. Zhu, Z. Jia, J. S. Strobl, M. Ehrich, H. P. Misra, and Y. Li, "Potent induction of total cellular and mitochondrial antioxidants and phase 2 enzymes by cruciferous sulforaphane in rat aortic smooth muscle cells: cytoprotection against oxidative and electrophilic stress," *Cardiovascular Toxicology*, vol. 8, no. 3, pp. 115–125, 2008.
- [141] M.-Y. Song, E.-K. Kim, W.-S. Moon et al., "Sulforaphane protects against cytokine- and streptozotocin-induced β -cell damage by suppressing the NF- κ B pathway," *Toxicology and Applied Pharmacology*, vol. 235, no. 1, pp. 57–67, 2009.
- [142] S.-D. Cho, G. Li, H. Hu et al., "Involvement of c-Jun N-terminal kinase in G2/M arrest and caspase-mediated apoptosis induced by sulforaphane in DU145 prostate cancer cells," *Nutrition and Cancer*, vol. 52, no. 2, pp. 213–224, 2005.
- [143] A. T. Dinkova-Kostova, P. Talalay, J. Sharkey et al., "An exceptionally potent inducer of cytoprotective enzymes: elucidation of the structural features that determine inducer potency and reactivity with KEAP1," *The Journal of Biological Chemistry*, vol. 285, no. 44, pp. 33747–33755, 2010.
- [144] A. T. Dinkova-Kostova and P. Talalay, "NAD(P)H:quinone acceptor oxidoreductase 1 (NQO1), a multifunctional antioxidant enzyme and exceptionally versatile cytoprotector," *Archives of Biochemistry and Biophysics*, vol. 501, no. 1, pp. 116–123, 2010.
- [145] A. T. Dinkova-Kostova, J. W. Fahey, A. L. Benedict et al., "Dietary glucoraphanin-rich broccoli sprout extracts protect against UV radiation-induced skin carcinogenesis in SKH-1 hairless mice," *Photochemical & Photobiological Sciences*, vol. 9, no. 4, pp. 597–600, 2010.
- [146] J. W. Fahey and T. W. Kensler, "Role of dietary supplements/nutraceuticals in chemoprevention through induction of cytoprotective enzymes," *Chemical Research in Toxicology*, vol. 20, no. 4, pp. 572–576, 2007.
- [147] R. C. W. Ma, "Genetics of cardiovascular and renal complications in diabetes," *Journal of Diabetes Investigation*, vol. 7, no. 2, pp. 139–154, 2016.
- [148] C. Rask-Madsen and G. L. King, "Vascular complications of diabetes: mechanisms of injury and protective factors," *Cell Metabolism*, vol. 17, no. 1, pp. 20–33, 2013.
- [149] X.-L. Chen, G. Dodd, S. Thomas et al., "Activation of Nrf2/ARE pathway protects endothelial cells from oxidant injury and inhibits inflammatory gene expression," *American Journal of Physiology. Heart and Circulatory Physiology*, vol. 290, no. 5, pp. H1862–H1870, 2006.
- [150] M. Zakkar, K. Van der Heiden, L. A. Luong et al., "Activation of Nrf2 in endothelial cells protects arteries from exhibiting a proinflammatory state," *Arteriosclerosis, Thrombosis, and Vascular Biology*, vol. 29, no. 11, pp. 1851–1857, 2009.
- [151] X. He, "Nrf2 is critical in defense against high glucose-induced oxidative damage in cardiomyocytes," *Journal of Molecular and Cellular Cardiology*, vol. 46, no. 1, pp. 47–58, 2009.
- [152] S. C. Lu, "Regulation of glutathione synthesis," *Molecular Aspects of Medicine*, vol. 30, no. 1-2, pp. 42–59, 2009.
- [153] L. Yuan and N. Kaplowitz, "Glutathione in liver diseases and hepatotoxicity," *Molecular Aspects of Medicine*, vol. 30, no. 1-2, pp. 29–41, 2009.
- [154] S. K. Biswas and I. Rahman, "Environmental toxicity, redox signaling and lung inflammation: the role of glutathione," *Molecular Aspects of Medicine*, vol. 30, no. 1-2, pp. 60–76, 2009.
- [155] A. Csiszar, N. Labinskyy, R. Jimenez et al., "Anti-oxidative and anti-inflammatory vasoprotective effects of caloric restriction in aging: role of circulating factors and SIRT1," *Mechanisms of Ageing and Development*, vol. 130, no. 8, pp. 518–527, 2009.
- [156] H. J. Kim and A. E. Nel, "The role of phase II antioxidant enzymes in protecting memory T cells from spontaneous apoptosis in young and old mice," *The Journal of Immunology*, vol. 175, no. 5, pp. 2948–2959, 2005.
- [157] L. C. Bailey-Downs, M. Mitschelen, D. Sosnowska et al., "Liver-specific knockdown of IGF-1 decreases vascular oxidative stress resistance by impairing the Nrf2-dependent antioxidant response: a novel model of vascular aging," *The Journals of Gerontology: Series A*, vol. 67, no. 4, pp. 313–329, 2012.
- [158] T. W. Sedlak, L. G. Nucifora, M. Koga et al., "Sulforaphane augments glutathione and influences brain metabolites in human subjects: a clinical pilot study," *Molecular Neuropsychiatry*, vol. 3, no. 4, pp. 214–222, 2018.
- [159] R. K. Thimmulappa, H. Lee, T. Rangasamy et al., "Nrf2 is a critical regulator of the innate immune response and survival during experimental sepsis," *The Journal of Clinical Investigation*, vol. 116, no. 4, pp. 984–995, 2006.
- [160] A. Ceriello, N. Bortolotti, E. Falletti et al., "Total radical-trapping antioxidant parameter in NIDDM patients," *Diabetes Care*, vol. 20, no. 2, pp. 194–197, 1997.

- [161] S. K. Biswas, D. E. Newby, I. Rahman, and I. L. Megson, "Depressed glutathione synthesis precedes oxidative stress and atherogenesis in Apo-E^{-/-} mice," *Biochemical and Biophysical Research Communications*, vol. 338, no. 3, pp. 1368–1373, 2005.
- [162] E. U. Nwose, H. F. Jelinek, R. S. Richards, and P. G. Kerr, "Erythrocyte oxidative stress in clinical management of diabetes and its cardiovascular complications," *British Journal of Biomedical Science*, vol. 64, no. 1, pp. 35–43, 2007.
- [163] E. U. Nwose, H. F. Jelinek, R. S. Richards, and P. G. Kerr, "Changes in the erythrocyte glutathione concentration in the course of diabetes mellitus," *Redox Report*, vol. 11, no. 3, pp. 99–104, 2006.
- [164] A. Prasad, N. P. Andrews, F. A. Padder, M. Husain, and A. A. Quyyumi, "Glutathione reverses endothelial dysfunction and improves nitric oxide bioavailability," *Journal of the American College of Cardiology*, vol. 34, no. 2, pp. 507–514, 1999.
- [165] A. Witschi, S. Reddy, B. Stofer, and B. H. Lauterburg, "The systemic availability of oral glutathione," *European Journal of Clinical Pharmacology*, vol. 43, no. 6, pp. 667–669, 1992.
- [166] M. Brownlee, "Biochemistry and molecular cell biology of diabetic complications," *Nature*, vol. 414, no. 6865, pp. 813–820, 2001.
- [167] M. Liang and J. L. Pietrusz, "Thiol-related genes in diabetic complications: a novel protective role for endogenous thioredoxin 2," *Arteriosclerosis, Thrombosis, and Vascular Biology*, vol. 27, no. 1, pp. 77–83, 2007.
- [168] J. Yang and R. H. Liu, "Induction of phase II enzyme, quinone reductase, in murine hepatoma cells *in vitro* by grape extracts and selected phytochemicals," *Food Chemistry*, vol. 114, no. 3, pp. 898–904, 2009.
- [169] A. Holmgren and J. Lu, "Thioredoxin and thioredoxin reductase: current research with special reference to human disease," *Biochemical and Biophysical Research Communications*, vol. 396, no. 1, pp. 120–124, 2010.
- [170] T. Ebrahimian and R. M. Touyz, "Thioredoxin in vascular biology: role in hypertension," *Antioxidants & Redox Signaling*, vol. 10, no. 6, pp. 1127–1136, 2008.
- [171] L. Tao, E. Gao, N. S. Bryan et al., "Cardioprotective effects of thioredoxin in myocardial ischemia and the reperfusion role of S-nitrosation," *Proceedings of the National Academy of Sciences of the United States of America*, vol. 101, no. 31, pp. 11471–11476, 2004.
- [172] K. C. Das, Y. Lewis-Molock, and C. W. White, "Elevation of manganese superoxide dismutase gene expression by thioredoxin," *American Journal of Respiratory Cell and Molecular Biology*, vol. 17, no. 6, pp. 713–726, 1997.
- [173] A. T. Dinkova-Kostova and P. Talalay, "Persuasive evidence that quinone reductase type I (DT diaphorase) protects cells against the toxicity of electrophiles and reactive forms of oxygen," *Free Radical Biology and Medicine*, vol. 29, no. 3–4, pp. 231–240, 2000.
- [174] Y. Yuan, L. Ji, L. Luo et al., "Quinone reductase (QR) inducers from *Andrographis paniculata* and identification of molecular target of andrographolide," *Fitoterapia*, vol. 83, no. 8, pp. 1506–1513, 2012.
- [175] A. T. Dinkova-Kostova, J. W. Fahey, K. L. Wade et al., "Induction of the phase 2 response in mouse and human skin by sulforaphane-containing broccoli sprout extracts," *Cancer Epidemiology, Biomarkers & Prevention*, vol. 16, no. 4, pp. 847–851, 2007.
- [176] D. Siegel, "NAD(P)H:quinone oxidoreductase 1: role as a superoxide scavenger," *Molecular Pharmacology*, vol. 65, no. 5, pp. 1238–1247, 2004.
- [177] E. L. Cavalieri and E. G. Rogan, "Depurinating estrogen–DNA adducts in the etiology and prevention of breast and other human cancers," *Future Oncology*, vol. 6, no. 1, pp. 75–91, 2010.
- [178] E. L. Cavalieri, D. E. Stack, P. D. Devanesan et al., "Molecular origin of cancer: Catechol estrogen-3,4-quinones as endogenous tumor initiators," *Proceedings of the National Academy of Sciences of the United States of America*, vol. 94, no. 20, pp. 10937–10942, 1997.
- [179] N. K. Idriss, A. D. Blann, and G. Y. Lip, "Hemoxygenase-1 in cardiovascular disease," *Journal of the American College of Cardiology*, vol. 52, no. 12, pp. 971–978, 2008.
- [180] M. He, R. C. Siow, D. Sugden, L. Gao, X. Cheng, and G. E. Mann, "Induction of HO-1 and redox signaling in endothelial cells by advanced glycation end products: A role for Nrf2 in vascular protection in diabetes," *Nutrition, Metabolism, and Cardiovascular Diseases*, vol. 21, no. 4, pp. 277–285, 2011.
- [181] S. A. Rushworth and M. A. O'Connell, "Haem oxygenase-1 in inflammation," *Biochemical Society Transactions*, vol. 32, no. 6, pp. 1093–1094, 2004.
- [182] A. Adaikalakoteswari, M. Balasubramanyam, M. Rema, and V. Mohan, "Differential gene expression of NADPH oxidase (p22phox) and hemoxygenase-1 in patients with Type 2 diabetes and microangiopathy," *Diabetic Medicine*, vol. 23, no. 6, pp. 666–674, 2006.
- [183] S. D. Yan, A. M. Schmidt, G. M. Anderson et al., "Enhanced cellular oxidant stress by the interaction of advanced glycation end products with their receptors/binding proteins," *Journal of Biological Chemistry*, vol. 269, no. 13, pp. 9889–9897, 1994.
- [184] R. A. Cohen and X. Tong, "Vascular oxidative stress: the common link in hypertensive and diabetic vascular disease," *Journal of Cardiovascular Pharmacology*, vol. 55, no. 4, pp. 308–316, 2010.
- [185] H. J. Prochaska, M. J. De Long, and P. Talalay, "On the mechanisms of induction of cancer-protective enzymes: a unifying proposal," *Proceedings of the National Academy of Sciences of the United States of America*, vol. 82, no. 23, pp. 8232–8236, 1985.
- [186] H. Raunio, K. Husgafvel-Pursiainen, S. Anttila, E. Hietanen, A. Hirvonen, and O. Pelkonen, "Diagnosis of polymorphisms in carcinogen-activating and inactivating enzymes and cancer susceptibility—a review," *Gene*, vol. 159, no. 1, pp. 113–121, 1995.
- [187] D. Liska, "Detoxification and biotransformational imbalances," *Explore*, vol. 2, no. 2, pp. 122–140, 2006.
- [188] S. Barcelo, J. M. Gardiner, A. Gescher, and J. K. Chipman, "CYP2E1-mediated mechanism of anti-genotoxicity of the broccoli constituent sulforaphane," *Carcinogenesis*, vol. 17, no. 2, pp. 277–282, 1996.
- [189] G. H. Posner, C. G. Cho, J. V. Green, Y. Zhang, and P. Talalay, "Design and synthesis of bifunctional isothiocyanate analogs of sulforaphane: correlation between structure and potency as inducers of anticarcinogenic detoxication enzymes," *Journal of Medicinal Chemistry*, vol. 37, no. 1, pp. 170–176, 1994.
- [190] L. B. von Weyarn, J. A. Chun, and P. F. Hollenberg, "Effects of benzyl and phenethyl isothiocyanate on P450s 2A6 and

- 2A13: potential for chemoprevention in smokers," *Carcinogenesis*, vol. 27, no. 4, pp. 782–790, 2006.
- [191] K. Skupinska, I. Misiewicz-Krzeminska, R. Stypulkowski, K. Lubelska, and T. Kasprzycka-Guttman, "Sulforaphane and its analogues inhibit CYP1A1 and CYP1A2 activity induced by benzo[a]pyrene," *Journal of Biochemical and Molecular Toxicology*, vol. 23, no. 1, pp. 18–28, 2009.
- [192] J. J. McElwee, E. Schuster, E. Blanc et al., "Evolutionary conservation of regulated longevity assurance mechanisms," *Genome Biology*, vol. 8, no. 7, article R132, 2007.
- [193] C. W. Nho and E. Jeffery, "The synergistic upregulation of phase II detoxification enzymes by glucosinolate breakdown products in cruciferous vegetables," *Toxicology and Applied Pharmacology*, vol. 174, no. 2, pp. 146–152, 2001.
- [194] R. H. Dashwood, "Indole-3-carbinol: anticarcinogen or tumor promoter in brassica vegetables?," *Chemico-Biological Interactions*, vol. 110, no. 1-2, pp. 1–5, 1998.
- [195] B. M. Lee and K. K. Park, "Beneficial and adverse effects of chemopreventive agents," *Mutation Research/Fundamental and Molecular Mechanisms of Mutagenesis*, vol. 523-524, pp. 265–278, 2003.
- [196] D. J. Kim, B. S. Han, B. Ahn et al., "Enhancement by indole-3-carbinol of liver and thyroid gland neoplastic development in a rat medium-term multiorgan carcinogenesis model," *Carcinogenesis*, vol. 18, no. 2, pp. 377–381, 1997.
- [197] S. T. Smale, "Hierarchies of NF- κ B target-gene regulation," *Nature Immunology*, vol. 12, no. 8, pp. 689–694, 2011.
- [198] C. S. Duckett, N. D. Perkins, T. F. Kowalik et al., "Dimerization of NF- κ B2 with RelA(p65) regulates DNA binding, transcriptional activation, and inhibition by an I kappa B-alpha (MAD-3)," *Molecular and Cellular Biology*, vol. 13, no. 3, pp. 1315–1322, 1993.
- [199] A. Oeckinghaus and S. Ghosh, "The NF- κ B family of transcription factors and its regulation," *Cold Spring Harbor Perspectives in Biology*, vol. 1, no. 4, article a000034, 2009.
- [200] H. Sakurai, S. Suzuki, N. Kawasaki et al., "Tumor necrosis factor- α -induced IKK phosphorylation of NF- κ B p65 on serine 536 is mediated through the TRAF2, TRAF5, and TAK1 signaling pathway," *Journal of Biological Chemistry*, vol. 278, no. 38, pp. 36916–36923, 2003.
- [201] Y. Goto and H. Kiyono, "Epithelial barrier: an interface for the cross-communication between gut flora and immune system," *Immunological Reviews*, vol. 245, no. 1, pp. 147–163, 2012.
- [202] X. L. Chen, G. Dodd, and C. Kunsch, "Sulforaphane inhibits TNF- α -induced activation of p38 MAP kinase and VCAM-1 and MCP-1 expression in endothelial cells," *Inflammation Research*, vol. 58, no. 8, pp. 513–521, 2009.
- [203] E. Heiss, C. Herhaus, K. Klimo, H. Bartsch, and C. Gerhauer, "Nuclear factor κ B is a molecular target for sulforaphane-mediated anti-inflammatory mechanisms," *Journal of Biological Chemistry*, vol. 276, no. 34, pp. 32008–32015, 2001.
- [204] S. Mukherjee, S. Karmakar, and S. P. Babu, "TLR2 and TLR4 mediated host immune responses in major infectious diseases: a review," *The Brazilian Journal of Infectious Diseases*, vol. 20, no. 2, pp. 193–204, 2016.
- [205] Y. Ben-Neriah and M. Karin, "Inflammation meets cancer, with NF- κ B as the matchmaker," *Nature Immunology*, vol. 12, no. 8, pp. 715–723, 2011.
- [206] M. Yu, H. Li, Q. Liu et al., "Nuclear factor p65 interacts with Keap1 to repress the Nrf2-ARE pathway," *Cellular Signalling*, vol. 23, no. 5, pp. 883–892, 2011.
- [207] S. Tao, L. Zhu, P. Lee et al., "Negative control of TLR3 signaling by TICAM1 down-regulation," *American Journal of Respiratory Cell and Molecular Biology*, vol. 46, no. 5, pp. 660–667, 2012.
- [208] S. M. Rahman, R. C. Janssen, M. Choudhury et al., "CCAA T/enhancer-binding protein β (C/EBP β) expression regulates dietary-induced inflammation in macrophages and adipose tissue in mice," *Journal of Biological Chemistry*, vol. 287, no. 41, pp. 34349–34360, 2012.
- [209] M. T. Lopez-Chillon, C. Carazo-Diaz, D. Prieto-Merino, P. Zafrilla, D. A. Moreno, and D. Villano, "Effects of long-term consumption of broccoli sprouts on inflammatory markers in overweight subjects," *Clinical Nutrition*, vol. 38, no. 2, pp. 745–752, 2018.
- [210] P. Mirmiran, Z. Bahadoran, F. Hosseinpanah, A. Keyzad, and F. Azizi, "Effects of broccoli sprout with high sulforaphane concentration on inflammatory markers in type 2 diabetic patients: a randomized double-blind placebo-controlled clinical trial," *Journal of Functional Foods*, vol. 4, no. 4, pp. 837–841, 2012.
- [211] S. H. Hu, J. C. Wang, H. F. Kung, J. T. Wang, W. L. Lee, and Y. H. Yang, "Antimicrobial effect of extracts of cruciferous vegetables," *The Kaohsiung Journal of Medical Sciences*, vol. 20, no. 12, pp. 591–599, 2004.
- [212] M. E. Martin and J. V. Solnick, "The gastric microbial community, *Helicobacter pylori* colonization, and disease," *Gut Microbes*, vol. 5, no. 3, pp. 345–350, 2014.
- [213] N. Wakabayashi, A. T. Dinkova-Kostova, W. D. Holtzclaw et al., "Protection against electrophile and oxidant stress by induction of the phase 2 response: fate of cysteines of the Keap1 sensor modified by inducers," *Proceedings of the National Academy of Sciences of the United States of America*, vol. 101, no. 7, pp. 2040–2045, 2004.
- [214] G. J. Fuchs, H. Mahmud, L. Davidsson et al., "*Helicobacter pylori* infection, iron absorption, and gastric acid secretion in Bangladeshi children," *The American Journal of Clinical Nutrition*, vol. 80, no. 1, pp. 149–153, 2004.
- [215] A. Yanaka, "Role of NRF2 in protection of the gastrointestinal tract against oxidative stress," *Journal of Clinical Biochemistry and Nutrition*, vol. 63, no. 1, pp. 18–25, 2018.
- [216] A. Yanaka, "Sulforaphane enhances protection and repair of gastric mucosa against oxidative stress in vitro, and demonstrates anti-inflammatory effects on *helicobacter pylori* infected gastric mucosae in mice and human subjects," *Current Pharmaceutical Design*, vol. 17, no. 16, pp. 1532–1540, 2011.
- [217] A. Yanaka, J. W. Fahey, A. Fukumoto et al., "Dietary sulforaphane-rich broccoli sprouts reduce colonization and attenuate gastritis in *Helicobacter pylori*-infected mice and humans," *Cancer Prevention Research*, vol. 2, no. 4, pp. 353–360, 2009.
- [218] M. V. Galan, A. A. Kishan, and A. L. Silverman, "Oral broccoli sprouts for the treatment of *Helicobacter pylori* infection: a preliminary report," *Digestive Diseases and Sciences*, vol. 49, no. 7-8, pp. 1088–1090, 2004.
- [219] L. V. Modolo, A. X. de Souza, L. P. Horta, D. P. Araujo, and A. de Fátima, "An overview on the potential of natural products as ureases inhibitors: a review," *Journal of Advanced Research*, vol. 6, no. 1, pp. 35–44, 2015.
- [220] J. W. Fahey, K. K. Stephenson, K. L. Wade, and P. Talalay, "Urease from *Helicobacter pylori* is inactivated by sulforaphane and other isothiocyanates," *Biochemical and*

- Biophysical Research Communications*, vol. 435, no. 1, pp. 1–7, 2013.
- [221] Y. F. Rego, M. P. Queiroz, T. O. Brito et al., “A review on the development of urease inhibitors as antimicrobial agents against pathogenic bacteria,” *Journal of Advanced Research*, vol. 13, pp. 69–100, 2018.
- [222] K. Kappaun, A. R. Piovesan, C. R. Carlini, and R. Ligabue-Braun, “Ureases: historical aspects, catalytic, and non-catalytic properties – a review,” *Journal of Advanced Research*, vol. 13, pp. 3–17, 2018.
- [223] A. Auron and P. D. Brophy, “Hyperammonemia in review: pathophysiology, diagnosis, and treatment,” *Pediatric Nephrology*, vol. 27, no. 2, pp. 207–222, 2012.
- [224] E. M. Pferschy-Wenzig and R. Bauer, “The relevance of pharmacognosy in pharmacological research on herbal medicinal products,” *Epilepsy & behavior*, vol. 52, Part B, pp. 344–362, 2015.
- [225] M. J. Rein, M. Renouf, C. Cruz-Hernandez, L. Actis-Goretta, S. K. Thakkar, and M. da Silva Pinto, “Bioavailability of bioactive food compounds: a challenging journey to bioefficacy,” *British Journal of Clinical Pharmacology*, vol. 75, no. 3, pp. 588–602, 2013.
- [226] R. H. Brown, C. Reynolds, A. Brooker, P. Talalay, and J. W. Fahey, “Sulforaphane improves the bronchoprotective response in asthmatics through Nrf2-mediated gene pathways,” *Respiratory Research*, vol. 16, article 106, 2015.
- [227] A. S. Axelsson, E. Tubbs, B. Mecham et al., “Sulforaphane reduces hepatic glucose production and improves glucose control in patients with type 2 diabetes,” *Science Translational Medicine*, vol. 9, no. 394, article eaah4477, 2017.
- [228] K. Singh, S. L. Connors, E. A. Macklin et al., “Sulforaphane treatment of autism spectrum disorder (ASD),” *Proceedings of the National Academy of Sciences of the United States of America*, vol. 111, no. 43, pp. 15550–15555, 2014.
- [229] B. G. Cipolla, E. Mandron, J. M. Lefort et al., “Effect of sulforaphane in men with biochemical recurrence after radical prostatectomy,” *Cancer Prevention Research*, vol. 8, no. 8, pp. 712–719, 2015.
- [230] D. Heber, Z. Li, M. Garcia-Lloret et al., “Sulforaphane-rich broccoli sprout extract attenuates nasal allergic response to diesel exhaust particles,” *Food & Function*, vol. 5, no. 1, pp. 35–41, 2014.
- [231] P. A. Egner, J. G. Chen, J. B. Wang et al., “Bioavailability of sulforaphane from two broccoli sprout beverages: results of a short-term, cross-over clinical trial in Qidong, China,” *Cancer Prevention Research*, vol. 4, no. 3, pp. 384–395, 2011.
- [232] Z. Bahadoran, P. Mirmiran, F. Hosseinpahan, A. Rajab, G. Asghari, and F. Azizi, “Broccoli sprouts powder could improve serum triglyceride and oxidized LDL/LDL-cholesterol ratio in type 2 diabetic patients: a randomized double-blind placebo-controlled clinical trial,” *Diabetes Research and Clinical Practice*, vol. 96, no. 3, pp. 348–354, 2012.
- [233] Z. Bahadoran, M. Tohidi, P. Nazeri, M. Mehran, F. Azizi, and P. Mirmiran, “Effect of broccoli sprouts on insulin resistance in type 2 diabetic patients: a randomized double-blind clinical trial,” *International Journal of Food Sciences and Nutrition*, vol. 63, no. 7, pp. 767–771, 2012.
- [234] L. L. Atwell, A. Hsu, C. P. Wong et al., “Absorption and chemopreventive targets of sulforaphane in humans following consumption of broccoli sprouts or a myrosinase-treated broccoli sprout extract,” *Molecular Nutrition & Food Research*, vol. 59, no. 3, pp. 424–433, 2015.
- [235] Y. W. Chang, J. Y. Jang, Y. H. Kim, J. W. Kim, and J. J. Shim, “The effects of broccoli sprout extract containing sulforaphane on lipid peroxidation and *Helicobacter pylori* infection in the gastric mucosa,” *Gut and Liver*, vol. 9, no. 4, pp. 486–493, 2015.
- [236] C. A. Houghton, R. G. Fassett, and J. S. Coombes, “Sulforaphane: translational research from laboratory bench to clinic,” *Nutrition Reviews*, vol. 71, no. 11, pp. 709–726, 2013.
- [237] J. W. Fahey, S. L. Wehage, W. D. Holtzclaw et al., “Protection of humans by plant glucosinolates: efficiency of conversion of glucosinolates to isothiocyanates by the gastrointestinal microflora,” *Cancer Prevention Research*, vol. 5, no. 4, pp. 603–611, 2012.
- [238] T. A. Shapiro, J. W. Fahey, A. T. Dinkova-Kostova et al., “Safety, tolerance, and metabolism of broccoli sprout glucosinolates and isothiocyanates: a clinical phase I study,” *Nutrition and Cancer*, vol. 55, no. 1, pp. 53–62, 2006.
- [239] X. Jiang, Y. Liu, L. Ma et al., “Chemopreventive activity of sulforaphane,” *Drug Design, Development and Therapy*, vol. 12, pp. 2905–2913, 2018.
- [240] R. J. DeBerardinis and N. S. Chandel, “Fundamentals of cancer metabolism,” *Science Advances*, vol. 2, no. 5, article e1600200, 2016.
- [241] H. M. Leinonen, E. Kansanen, P. Polonen, M. Heinaniemi, and A. L. Levenon, “Dysregulation of the Keap1–Nrf2 pathway in cancer,” *Biochemical Society Transactions*, vol. 43, no. 4, pp. 645–649, 2015.
- [242] H. M. Leinonen, E. Kansanen, P. Pölönen, M. Heinaniemi, and A.-L. Levenon, “Role of the Keap1–Nrf2 pathway in cancer,” *Advances in Cancer Research*, vol. 122, pp. 281–320, 2014.
- [243] E. Kansanen, S. M. Kuosmanen, H. Leinonen, and A. L. Levenon, “The Keap1–Nrf2 pathway: mechanisms of activation and dysregulation in cancer,” *Redox Biology*, vol. 1, pp. 45–49, 2013.
- [244] J. D. Hayes and A. T. Dinkova-Kostova, “The Nrf2 regulatory network provides an interface between redox and intermediary metabolism,” *Trends in Biochemical Sciences*, vol. 39, no. 4, pp. 199–218, 2014.
- [245] J. M. Maher, M. Z. Dieter, L. M. Aleksunes et al., “Oxidative and electrophilic stress induces multidrug resistance-associated protein transporters via the nuclear factor-E2-related factor-2 transcriptional pathway,” *Hepatology*, vol. 46, no. 5, pp. 1597–1610, 2007.
- [246] “Nobel-Laureate,” 1962, <https://www.nobelprize.org/prizes/medicine/1962/summary/>.
- [247] J. Watson, “Oxidants, antioxidants and the current incurability of metastatic cancers,” *Open biology*, vol. 3, no. 1, article 120144, 2013.
- [248] D. Seely and S. Verma, “The Oncology Association of Naturopathic Physicians principles of care guidelines,” *Current Oncology*, vol. 26, no. 1, pp. 8–9, 2019.
- [249] T. Forster, V. Rausch, Y. Zhang et al., “Sulforaphane counteracts aggressiveness of pancreatic cancer driven by dysregulated Cx43-mediated gap junctional intercellular communication,” *Oncotarget*, vol. 5, no. 6, pp. 1621–1634, 2014.
- [250] W. Wang, Y. He, G. Yu et al., “Sulforaphane protects the liver against CdSe quantum dot-induced cytotoxicity,” *PLoS One*, vol. 10, no. 9, article e0138771, 2015.

- [251] X. Wang, Y. Li, Y. Dai et al., "Sulforaphane improves chemotherapy efficacy by targeting cancer stem cell-like properties via the miR-124/IL-6R/STAT3 axis," *Scientific Reports*, vol. 6, article 36796, 2016.
- [252] C. Kerr, G. Adhikary, D. Grun, N. George, and R. L. Eckert, "Combination cisplatin and sulforaphane treatment reduces proliferation, invasion, and tumor formation in epidermal squamous cell carcinoma," *Molecular Carcinogenesis*, vol. 57, no. 1, pp. 3–11, 2018.
- [253] N. Khurana, H. Kim, P. K. Chandra et al., "Multimodal actions of the phytochemical sulforaphane suppress both AR and AR-V7 in 22Rv1 cells: advocating a potent pharmaceutical combination against castration-resistant prostate cancer," *Oncology Reports*, vol. 38, no. 5, pp. 2774–2786, 2017.
- [254] C. Bose, S. Awasthi, R. Sharma et al., "Sulforaphane potentiates anticancer effects of doxorubicin and attenuates its cardiotoxicity in a breast cancer model," *PLoS One*, vol. 13, no. 3, article e0193918, 2018.
- [255] A. Thakkar, D. Sutaria, B. K. Grandhi, J. Wang, and S. Prabhu, "The molecular mechanism of action of aspirin, curcumin and sulforaphane combinations in the chemoprevention of pancreatic cancer," *Oncology Reports*, vol. 29, no. 4, pp. 1671–1677, 2013.
- [256] M. F. Ullah, "Sulforaphane (SFN): an isothiocyanate in a cancer chemoprevention paradigm," *Medicines*, vol. 2, no. 3, pp. 141–156, 2015.
- [257] A. F. Abdull Razis and N. M. Noor, "Cruciferous vegetables: dietary phytochemicals for cancer prevention," *Asian Pacific Journal of Cancer Prevention*, vol. 14, no. 3, pp. 1565–1570, 2013.
- [258] X. Zhang, X. O. Shu, Y. B. Xiang et al., "Cruciferous vegetable consumption is associated with a reduced risk of total and cardiovascular disease mortality," *The American Journal of Clinical Nutrition*, vol. 94, no. 1, pp. 240–246, 2011.
- [259] A. Leone, G. Diorio, W. Sexton et al., "Sulforaphane for the chemoprevention of bladder cancer: molecular mechanism targeted approach," *Oncotarget*, vol. 8, no. 21, pp. 35412–35424, 2017.
- [260] A. Lan, W. Li, Y. Liu et al., "Chemoprevention of oxidative stress-associated oral carcinogenesis by sulforaphane depends on NRF2 and the isothiocyanate moiety," *Oncotarget*, vol. 7, no. 33, pp. 53502–53514, 2016.
- [261] L. Yang, D. L. Palliyaguru, and T. W. Kensler, "Frugal chemoprevention: targeting Nrf2 with foods rich in sulforaphane," *Seminars in Oncology*, vol. 43, no. 1, pp. 146–153, 2016.
- [262] A. I. Amjad, R. A. Parikh, L. J. Appleman, E. R. Hahm, K. Singh, and S. V. Singh, "Broccoli-derived sulforaphane and chemoprevention of prostate cancer: from bench to bedside," *Current Pharmacology Reports*, vol. 1, no. 6, pp. 382–390, 2015.
- [263] S. M. Tortorella, S. G. Royce, P. V. Licciardi, and T. C. Karagiannis, "Dietary sulforaphane in cancer chemoprevention: the role of epigenetic regulation and HDAC inhibition," *Antioxidants & Redox Signaling*, vol. 22, no. 16, pp. 1382–1424, 2015.
- [264] A. Qazi, J. Pal, M. Maitah et al., "Anticancer activity of a broccoli derivative, sulforaphane, in Barrett adenocarcinoma: potential use in chemoprevention and as adjuvant in chemotherapy," *Translational Oncology*, vol. 3, no. 6, pp. 389–399, 2010.



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